

Distinct patterns in the diurnal and seasonal variability in four components of soil respiration in a temperate forest under free-air CO₂ enrichment

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Received: 1 March 2011 – Published in Biogeosciences Discuss.: 18 March 2011

Revised: 10 October 2011 – Accepted: 21 October 2011 – Published: 31 October 2011

Abstract. Soil respiration (R_S) is a major flux in the global carbon (C) cycle. Responses of R_S to changing environmental conditions may exert a strong control on the residence time of C in terrestrial ecosystems and in turn influence the atmospheric concentration of greenhouse gases. Soil respiration consists of several components oxidizing soil C from different pools, age and chemistry. The mechanisms underlying the temporal variability of R_S components are poorly understood. In this study, we used the long-term whole-ecosystem ¹³C tracer at the Duke Forest Free Air CO₂ Enrichment site to separate forest R_S into its autotrophic (R_R) and heterotrophic components (R_H). The contribution of R_H to R_S was further partitioned into litter decomposition (R_L), and decomposition of soil organic matter (R_{SOM}) of two age classes – up to 8 yr old and SOM older than 8 yr. Soil respiration was generally dominated by R_{SOM} during the growing season (44% of daytime R_S), especially at night. The contribution of heterotrophic respiration (R_{SOM} and R_L) to R_S was not constant, indicating that the seasonal variability in R_R alone cannot explain seasonal variation in R_S . Although there was no diurnal variability in R_S , there were significant compensatory differences in the contribution of individual R_S components to daytime and nighttime rates. The average contribution of R_{SOM} to R_S was greater at night (54%) than during the day (44%). The average contribution of R_R to total R_S was ~30% during the day and ~34% during the night. In contrast, R_L constituted 26% of R_S during the day and only 12% at night. About 95% of the decomposition of soil C older than 8 yr (R_{pre-tr}) originated from R_{SOM} and

showed more pronounced and consistent diurnal variability than any other R_S component; nighttime rates were on average 29% higher than daytime rates. In contrast, the decomposition of more recent, post-treatment C (R_{pre-tr}) did not vary diurnally. None of the diurnal variations in components of R_H could be explained by only temperature and moisture variations. Our results indicate that the variation observed in the components of R_S is the result of complex interaction between dominant biotic controls (e.g. plant activity, mineralization kinetics, competition for substrates) over abiotic controls (temperature, moisture). The interactions and controls among roots and other soil organisms that utilize C of different chemistry, accessibility and ages, results in the overall soil CO₂ efflux. Therefore understanding the controls on the components of R_S is necessary to elucidate the influence of ecosystem respiration on atmospheric C-pools at different time scales.

1 Introduction

Terrestrial ecosystems exchange large amounts of C with the atmosphere through the processes of photosynthesis and ecosystem respiration (R_E). Annually, the difference between these large fluxes determines the extent of C storage in the terrestrial biosphere and small imbalances between these fluxes can lead to substantial variation in atmospheric CO₂ concentration. The role of ecosystems as a long-term sink or source for atmospheric C thus depends on the effects and feedbacks of changing environmental conditions on photosynthesis and the components of R_E . The potential responses of R_E to environmental change are less clear than those of photosynthesis (Gonzalez-Meler et al., 2004; DeLucia et al.,



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2007), but are of fundamental importance in determining the residence time of C in terrestrial ecosystems. Improved understanding of the biotic and abiotic mechanisms controlling C release from terrestrial ecosystems, and the time scales at which these mechanisms operate, is necessary before the future role of the terrestrial biosphere in the global C cycle can be predicted.

Ecosystem respiration is often dominated by soil respiration (R_S), which can constitute 50–80 % of the total C emitted from ecosystems to the atmosphere annually (Raich et al., 2002; Davidson and Janssens, 2006; Davidson et al., 2006). Soil respiration results from a complex network of oxidation processes, involving different substrates of various ages and carried out by different organisms at different temporal and spatial scales (Taneva et al., 2006). Soil respiration includes respiration by live roots, root-associated microorganisms, and microbial decomposition of root exudates (collectively referred to as root/rhizosphere respiration, R_R), as well as from heterotrophic respiration (R_H) associated with the decomposition of root and leaf litter, and other soil organic matter (SOM) pools of different ages. Ecosystem exposure to elevated $[CO_2]$ or high temperature has been shown to lead to enhanced R_S rates only initially (King et al., 2004; Bernhardt et al., 2006; Melillo et al., 2002). It remains unclear whether changes in R_S of ecosystems exposed to elevated CO_2 or warming are the result of increased R_R , R_H , or a combination of both (Gonzalez-Meler and Taneva, 2005; Subke et al., 2006; Bradford et al., 2008).

Because individual components of R_S return soil carbon of different nature and age back to the atmosphere, a shift in their relative contributions to total R_S with environmental changes, will impact the residence time of soil C and, therefore, atmospheric CO_2 concentration levels. For instance, atmospheric CO_2 enrichment may cause increases in below-ground allocation (Matamala and Schlesinger, 2000; Norby et al., 2002) leading to increased total R_R rates. Increases in R_S rates caused solely by a photosynthesis-driven direct enhancement of R_R may have little consequence to SOM pool changes and atmospheric CO_2 concentration. Greater soil C inputs under elevated $[CO_2]$ may also increase substrate availability to soil microorganisms and lead to higher R_H rates (Hamilton et al., 2002; Makiranta et al., 2008; Wei et al., 2010). Heterotrophic respiration returns older soil C to the atmosphere and changes in both the sources and rates of R_H with environmental conditions (e.g. elevated $[CO_2]$, plant activity, altered soil moisture and/or temperature) could substantially affect the C sink capacity and turnover of soil C, with the potential to affect atmospheric $[CO_2]$.

Partitioning R_S into its components is inherently difficult and a variety of methods have been applied to the separation of R_R from R_H (Hanson et al., 2000; Subke et al., 2006). The average contribution of R_R to total R_S in temperate forests has been estimated to be ~45 %, with a range of 10 to 90 % (Hanson et al., 2000; Bond-Lamberty et al., 2004). The proportion of R_R has been shown to be related to annual

R_S rates and may not be constant across temporal or spatial scales (Bond-Lamberty et al., 2004; Subke et al., 2006; Kuzyakov and Gavrichkova, 2010), challenging the use of a single annual value for R_R/R_S in terrestrial C cycle models. An emerging pattern from R_E partitioning studies is that photosynthesis exerts a strong influence on R_S on diel and seasonal time scales (Högberg et al., 2001; Bowling et al., 2002; Tang et al., 2005; Kuzyakov and Gavrichkova, 2010). Trueman and Gonzalez-Meler (2005) showed that the rates of oxidation of soil pools that contained C older than 4 yr were highly influenced by changes in plant activity. These observations suggest that there are complex interactive effects between R_S components that may operate at different time scales, involving several soil C pools that may differ in chemical composition and soil residence time (Heath et al., 2005). The interactive effects of biotic and abiotic variables on R_S and its components have not been elucidated.

Temperature- and moisture-dependent models are widely used for predicting the response of terrestrial ecosystems to changing environmental conditions (Lloyd and Taylor, 1994; Reichstein et al., 2003; Luo, 2007). Individual components of R_S , however, can often be independently affected by other abiotic or biotic variables, as well as by their interactions (Trueman and Gonzalez-Meler, 2005; Kuzyakov and Gavrichkova, 2010). A significant amount of photosynthetic carbon is returned to the atmosphere through R_R within days of assimilation (Ekblad and Hogberg, 2001; Bowling et al., 2002; Trueman and Gonzalez-Meler, 2005; Taneva et al., 2006; Carbone et al., 2007; Mencuccini and Hölttä, 2010; Kuzyakov and Gavrichkova, 2010), highlighting the importance of photosynthesis in influencing R_S rates. Enhanced plant activity may also lead to changes in the decomposition rate of older SOM through “priming”, if they result in changes in the size of the SOM pool (Kuzyakov, 2002; Subke et al., 2004). Biotic controls on the rate of the components of R_S can also be confounded with the temperature- and moisture-dependent functions often used to describe variations in R_S at seasonal time scales, potentially leading to limitations in our mechanistic predictions of ecosystem C budgets (Liu et al., 2006).

In this study, we used litter removal and the long-term ^{13}C tracer at the Duke Forest Free Air CO_2 Enrichment (FACE) experiment (Chapel Hill, NC, USA) to partition growing season R_S into the contributions of root/rhizosphere respiration, litter decomposition, and decomposition of SOM. We also separated an older than 8 yr C pool based on the time at which elevated CO_2 exposure began (1996). Stable isotope labeling techniques have been used successfully to partition R_S into some of its components (e.g. Andrews et al., 1999; Matamala et al., 2003; Taneva et al., 2006) as isotopes provide a non-disruptive alternative to destructive methods for distinguishing the origin of soil-respired CO_2 . Our specific objectives were: (1) to determine the diel and seasonal variability of the components of R_S ; and (2) to understand how variations in R_S components affect observed rates of R_S .

2 Materials and methods

2.1 Site description

The Forest Atmosphere Carbon Transfer and Storage 1 (FACTS-1) research site is located in the Blackwood Division of the Duke Forest, near Chapel Hill, North Carolina, USA (35°58' N 79°05' W). The Free Air CO₂ Enrichment (FACE) experiment at FACTS-1 consists of six 30-m diameter plots in an intact *Pinus taeda* plantation. Of the six plots, three are fumigated with CO₂ to maintain atmospheric [CO₂] about 200 µl l⁻¹ above ambient levels (567 ± 4 µl l⁻¹ 1996–2004; K. Lewin and R. Nettles, personal communication, 2009). The other three control plots are fumigated with ambient air only (Hendrey et al., 1999). Continuous fumigation of all plots began on 27 August 1996, 15 yr after planting. CO₂ fumigation is switched off when temperatures are below 5 °C and when sustained wind speed exceeds 5 m s⁻¹ and since 2003 fumigation was limited to daytime only.

Although dominated by pines through natural succession, a number of hardwood species have become established in the understory (*Acer rubrum*, *Liquidambar styraciflua*, *Liriodendron tulipifera*, *Ulmus alata*, and *Cercis Canadensis*). Soils at the site are clay-rich, low fertility Ultic Alfisols, with a pH of about 5. Fine roots are found mostly in the upper 20 cm of the soil profile (Matamala and Schlesinger, 2000). Mean annual temperature is 15.5 °C and mean annual precipitation is 1140 mm.

2.2 Ecosystem ¹³C tracer

The CO₂ used in FACE experiments is usually depleted in ¹³C ($\delta^{13}\text{C} \approx -43.1 \pm 0.6\text{‰}$ vs. PDB, where $\delta^{13}\text{C} = [(R_{\text{sample}} - R_{\text{reference}})/R_{\text{reference}}] \cdot 1000$ and $R = {}^{13}\text{C}/{}^{12}\text{C}$). The CO₂ released in the elevated [CO₂] plots has a $\delta^{13}\text{C}$ of about -20 ‰. The isotopic shift caused by the ¹³C-depleted CO₂ continuously applied in the elevated [CO₂] plots allows for distinguishing C from plant and soil material produced before starting the experiment ($\delta^{13}\text{C}$ of -29.9 ± 0.2 ‰ and -27.6 ± 0.2 ‰ for needles and roots respectively) and plant material produced during the experiment after 1996 (41.8 ± 0.3 ‰ and -39.7 ± 0.8 ‰ for needles and roots, respectively; see Matamala et al., 2003; Taneva and Gonzalez-Meler, 2008, for examples). The ¹³C label has also slowly been incorporated into soil organic matter pools and in soil-respired CO₂ (Andrews et al., 1999; Schlesinger and Lichter, 2001; Taneva et al., 2006; Lichter et al., 2008; Taneva and Gonzalez-Meler, 2008). The different rate at which the ¹³C label is incorporated into respired-CO₂ of soils components (Taneva et al., 2006) allow for the separation of root-respired CO₂ from SOM-respired CO₂ (see below).

2.3 Growing season soil respiration and litter removal treatment

During the 2003 and 2004 growing seasons, soil respiration rates were measured with a field-portable infrared gas analyzer (IRGA; LiCor 6400-09, Lincoln, Nebraska, USA) at 12 PVC collars, randomly placed within each FACE plot inserted 3 cm into the mineral soil and open to rainfall and litterfall, except during measurements. In May 2004, four additional soil collars were installed in each FACE plot, where the litter layer was completely removed down to the mineral soil. A layer of inert fiber glass was placed over the soil in order to reproduce the CO₂ diffusivity and moisture content of the removed litter. Soil respiration rates were measured monthly during the growing season of the forest (May–October), both during the day (12:00–14:00 EST) and at night (22:00–00:00 EST). Measurements were made at the times previously determined to capture most of the diurnal variability in soil respiration rates. The six FACE plots were grouped into three blocks, each including one treatment and one control plot. The measurement time in each plot was ~1 h and, therefore, only one block was measured on a given day, in order to ensure time consistency of measurements. Measurements in all three blocks were carried out on days with comparable environmental conditions and were usually completed within 5–6 days.

2.4 Stable isotope analysis of soil-respired CO₂

During the 2003 and 2004 growing seasons, soil-respired CO₂ samples were collected monthly from collars with and without litter, both during the day and at night, within 24 h after soil respiration measurements were made (see above). Carbon dioxide gas samples were collected from a LiCor 6400-09 soil chamber into evacuated 120-ml glass flasks, after being passed through a magnesium perchlorate water trap (Still et al., 2003; Trueman and Gonzalez-Meler, 2005; Moore et al., 2008). The CO₂ concentration of each sample was measured at the time of sample collection. At least eight gas samples from collars containing litter or no litter layer were collected from each FACE plot at each sampling time. Samples were collected from different collars to avoid alterations of convective patterns of CO₂ from soil to air and other recognized problems when collecting soil surface fluxes for building keeling plots (Phillips and Greg, 2001; Trueman and Gonzalez-Meler, 2005; Bowling et al., 2008; Kayler et al., 2010). Different collars were used because previous trials made both in May of 2003 and 2004 showed that the constructed Keeling plots obtained from a single location or multiple nearby collar location were not different (see the Supplement, Still et al., 2003). Gas samples were collected at CO₂ concentrations that differed by at least 50 ppm from other samples. Samples were shipped to the University of Illinois at Chicago for stable isotope analysis. In the laboratory, soil-respired CO₂ samples were purified by cryogenic

Table 1. The seasonal average ^{13}C signature of respired CO_2 and bulk mass from roots, root-free soil organic matter, and litter from control and treatment plots at FACTS-1. The average values listed here were derived from several day and night field incubations (see methods) for each collar location and for each time soil respiration and keeling plots were made in 2004 (June through September). Average values are expressed in per mil \pm standard error ($n = 3$).

	Ambient [CO_2]		Elevated [CO_2]	
	Respired CO_2	Bulk Mass	Respired CO_2	Bulk Mass
Roots	-29.0 ± 0.5	-27.6 ± 0.4	-40.4 ± 1.0	-39.7 ± 0.5
Root-free SOM	-26.5 ± 0.1	-26.3 ± 0.3	-34.5 ± 0.6	-30.2 ± 0.1
Forest floor Litter	-28.7 ± 0.4	-27.9 ± 0.2	-37.9 ± 0.6	-39.6 ± 0.3

extraction before they were analyzed for their stable C isotope composition with a Finnegan Delta Plus XL (Bremen, Germany) isotope ratio mass spectrometer. The $\delta^{13}\text{C}$ of soil-respired CO_2 , in the absence of atmospheric air, was determined using Keeling Plot analyses (Pataki et al., 2003). The range in [CO_2] of samples used to construct Keeling Plots was at least 480 ppm. Keeling Plot regressions with an r^2 value of <0.90 were excluded from further analysis.

2.5 Isotopic composition of root-, SOM- and litter-respired CO_2

The isotopic composition of root-, SOM- and litter-respired CO_2 was measured (following Hymus et al., 2005 and Trueman and Gonzalez-Meler, 2005) and used as endmembers in order to partition the root, SOM and litter contributions to soil respiration. Throughout the growing season of 2004, soil cores (0–10 cm, 2 cm diameter) were collected from locations adjacent to collars after respiration and isotope measurements (keeling plots) were done. Top 10 cm were chosen as it contains more than 90 % of the fine root biomass (Matamala et al., 2003) and has the most changes in pore space soil CO_2 concentration (Taneva et al., 2006) and C mineralization levels (Lichter et al., 2008; Taneva and Gonzalez-Meler, 2008). For endmember determination by incubations, roots and leaf litter were removed from the soil immediately after collection. Live fine roots (<5 mm diameter) were further rinsed in distilled water of all attached soil. Soil was removed from the litter by hand. In order to determine the $\delta^{13}\text{C}$ of respired CO_2 , litter, roots, and root-free soil (SOM) collected from each FACE plot were incubated separately in the dark in custom-designed PVC chambers with screw caps (400-ml chambers for soil and litter incubations and 150-ml chambers for root incubation), following Hymus et al. (2005), Trueman and Gonzalez-Meler (2005) and Taneva and Gonzalez-Meler (2008). Incubations were done at soil (root, SOM) and forest floor temperature (litter) at the time of collection. The field incubation system consisted of a pump, a soda lime column placed before the incubation chamber, a desiccant column placed between the chamber and the glass flask, where the respired CO_2 was eventually collected, and

an IRGA (LiCor 6262, Lincoln, Nebraska, USA). All components of the incubation system were connected to each other with Bev-A-Line[®] tubing (1/4" outer diameter). Before sample incubation, the chamber, the 150-ml glass flask, and the line were flushed with CO_2 -free air by pumping dry ambient air through the soda lime column. An Infra Red Gas Analyzer was used to monitor the [CO_2] of the air in the incubation system and trapped sample. The air-tight chamber remained close with three-way valves (Swagelok, Solon, OH, USA) for an incubation period of 20–30 min, depending on respiration rate. After the incubation and prior to collecting the respired CO_2 from each sample, the incubation system was once again flushed with CO_2 -free air, bypassing the closed incubation chamber, to ensure the lines and flask were free of H_2O and CO_2 . Then, valves from the incubation chamber were opened and the CO_2 -free air carried the sample respired- CO_2 into the glass flask, with concentrations ranging from 400 to 1200 ppm. Flasks containing the dried gas samples were shipped to the University of Illinois at Chicago for analysis. These incubation experiments were also done at the ambient rings to account for the environmental variability in the isotopic composition of respired CO_2 that are independent from the addition of the post treatment isotope label. The $\delta^{13}\text{C}$ value of respired CO_2 from roots, litter, and root-free soil from each plot at each sampling time (see Table 1 for averages) was used in the partitioning of soil-respired CO_2 into its source components (see below).

2.6 Partitioning soil-respired CO_2 into its R_R , R_{SOM} and R_L components in the 2004 growing season

There is a large isotopic difference between the C that was fixed by the ecosystem after CO_2 fumigation started compared to the existing ecosystem C (see Sect. 2.7 for details). Also the isotope air label in elevated CO_2 plots was rapidly incorporated in soil CO_2 , soil respiration and new roots (days to months; Andrews et al., 1999; Matamala et al., 2003; Taneva et al., 2006), moderately incorporated in existing roots and litter (years; Matamala et al., 2003; Lichter et al., 2008) and slowly into SOM pools (decades; Lichter et al., 2008). However, at this site the isotopic composition of

static pools does not correspond to the isotopic composition of metabolically active pools (Taneva et al., 2006) and therefore the isotopic composition of respired CO₂ from roots, SOM, and forest floor litter need to be measured and used in partitioning mixing models. There is a large isotopic difference between the $\delta^{13}\text{C}$ of respired CO₂ from roots and SOM (>6 per mil), but the difference with forest floor litter is too small (within 3 per mil). The litter exclusion experiments enabled us to further partition soil-respired CO₂ into CO₂ originating from root/rhizosphere respiration (R_R), litter decomposition (R_L), and SOM decomposition (R_{SOM}). This is because in absence of litter, the $\delta^{13}\text{C}$ of respired CO₂ is related to contributions from only roots and SOM to R_S with large isotopic differences. Once the proportions of R_R and R_{SOM} are known, R_L can be derived from the $\delta^{13}\text{C}$ of R_S measured with litter. Using two mixing equations with two unknowns, the contribution of R_R , R_{SOM} , and R_L to total R_S can then be expressed as follows:

$$\delta^{13}\text{C}_{R_S\text{CO}_2} = a \cdot \delta^{13}\text{C}_{\text{root CO}_2} + b \cdot \delta^{13}\text{C}_{\text{SOM CO}_2} + (1 - (a + b)) \cdot \delta^{13}\text{C}_{\text{litter CO}_2}, \quad (1)$$

where the fraction of root-respired CO₂ (a) was determined from Eq. (4), b is the fraction of soil-respired CO₂ produced by SOM decomposition, and the remaining CO₂ in R_S , determined as $(1 - (a + b))$, represents CO₂ produced in forest floor litter decomposition.

Assuming that the ratio of R_R to R_{SOM} in the plots without litter is the same as that in plots with litter, the fractions of R_R and R_{SOM} in plots without litter (nl) can be expressed as follows:

$$\delta^{13}\text{C}_{\text{nl CO}_2} = (a/(a + b)) \cdot \delta^{13}\text{C}_{\text{root CO}_2} + (b/(a + b)) \cdot \delta^{13}\text{C}_{\text{SOM CO}_2}, \quad (2)$$

where a represents the fraction of root-respired CO₂ in R_S , b is the fraction of SOM decomposition in R_S (as in Eq. 3), $\delta^{13}\text{C}_{\text{nl CO}_2}$ is the $\delta^{13}\text{C}$ of CO₂ from collars with no litter (from Keeling Plot analyses), $\delta^{13}\text{C}_{\text{root}}$ is the $\delta^{13}\text{C}$ of root-respired CO₂ (from root incubations), and $\delta^{13}\text{C}_{\text{SOM}}$ is the $\delta^{13}\text{C}$ measured with litter- and root-free soil incubations (incubation methods described in Sect. 2.5).

To calculate the actual respiration rate each of these components contribute to total R_S , the fractional values of a , b , and $(1 - (a + b))$ calculated over the growing season were multiplied by the measured R_S rate for each of the 12 collars per plot at a given time of the growing season when measurements were made. Then the 12 R_S locations per plot were averaged for each FACE ring, the replication unit, before the treatment average was measured ($n = 3$). We report here the R_L rates calculated with the isotope method for consistency and because of statistical power as they were calculated from 12 locations per plot (as oppose to 4 locations per plot using the R_S and R_{nl} difference). Also R_L measured as a residual from the litter exclusion experiments based on 4 replicates introduce artifacts due to variable forest floor mass.

2.7 Determination of pre- and post-treatment C in R_S

Soil-respired CO₂ can be partitioned into C that was photosynthetically fixed since the beginning of CO₂ fumigation (referred to as “post-treatment” C) and C assimilated under ambient [CO₂] before fumigation started in September, 1996 (referred to as “pre-treatment” C). We used the following two end-member mixing equation:

$$\delta^{13}\text{C}_{R_S\text{CO}_2} = f \cdot \delta^{13}\text{C}_{\text{pre-tr}} + (1 - f) \cdot \delta^{13}\text{C}_{\text{post-tr}} \quad (3)$$

where $\delta^{13}\text{C}_{R_S\text{CO}_2}$ is the measured $\delta^{13}\text{C}$ of soil-respired CO₂ at a given time, $\delta^{13}\text{C}_{\text{post-tr}}$ is the end-member for post-treatment C, $\delta^{13}\text{C}_{\text{pre-tr}}$ is the end-member for pre-treatment C (usually separated by about 12 per mil) and f represents the fraction of pre-treatment C in soil CO₂ (Taneva et al., 2006). This partitioning was done for all collection times during the 2003 and 2004 growing seasons.

The $\delta^{13}\text{C}_{\text{pre-tr}}$ is determined by directly measuring $\delta^{13}\text{C}_{R_S\text{CO}_2}$ in the control plots of the experiment at time of measurements. This measured value incorporates respiration from both recalcitrant and labile soil C pools under ambient CO₂ conditions. The $\delta^{13}\text{C}$ of recalcitrant soil C pools has little or no seasonal variation (Balesdent and Mariotti, 1996) and therefore the $\delta^{13}\text{C}$ of respired-CO₂ from this pool will not vary. Therefore, any seasonal variability in $\delta^{13}\text{C}$ of soil-respired CO₂ in the control plots is due to differences in the signature of respired CO₂ from labile soil C pools (i.e. root/rhizosphere respiration), reflecting, for instance, seasonal fluctuations in photosynthetic discrimination. There are two tested conditions that allow us to use ambient signature of soil-respired CO₂ in the calculations: (1) the $\delta^{13}\text{C}$ of the atmosphere in the CO₂-enriched plots was changed by a constant value E at the beginning of the experiment, and (2) the photosynthetic discrimination against ^{13}C is very similar under ambient and elevated [CO₂] due to conserved C_i/C_a for concentrations ranges below $\sim 700 \mu\text{l l}^{-1}$ (Ellsworth, 1999; Katul et al., 2010) making the difference in $\delta^{13}\text{C}$ of new photosynthate in the control and treatment plots to approximate E . Therefore, the end-member for the $\delta^{13}\text{C}$ of soil-respired CO₂ in the enriched plots ($\delta^{13}\text{C}_{\text{post-tr}}$) can be derived by subtracting E from the measured $\delta^{13}\text{C}_{\text{pre-tr}}$ and Eq. (3) can be rearranged as follows:

$$f = (\delta^{13}\text{C}_{R_S\text{CO}_2} - \delta^{13}\text{C}_{\text{post-tr}}) / E \quad (4)$$

where E is measured to be $11.82 \pm 0.43 \text{‰}$ based on: (i) the 1996–2004 plot average [CO₂] and $\delta^{13}\text{C}$ of fumigation CO₂; (ii) change in isotopic composition of new leaf tissue between ambient and elevated [CO₂]; (iii) change in isotopic composition of new in-growth root tissue between ambient and elevated [CO₂]; (iv) the difference in the isotopic composition in root-respired CO₂ between ambient and elevated [CO₂]. This value of $\sim 12 \text{‰}$ has been widely applied for isotope mixing models at the site (e.g. Andrews et al., 1999; Schlesinger and Lichter, 2001; Matamala et al., 2003; Bernhardt et al., 2006; Lichter et al., 2008). These pre- and

post-treatment C partitioning were done for the growing seasons of 2003 and 2004.

For the growing season of 2004, we used pre-treatment C flux to further partition R_S . In 2004, pre-treatment CO_2 efflux originated from soil C pools that were at least 8 yr. At the study site, the mean residence times of root and forest floor C are about 4 and 2.5 yr, respectively (Matamala et al., 2003; Lichter et al., 2008). It is unlikely that substantial amounts of storage carbohydrates contributed to R_R after 8 yr. It is also unlikely that forest floor litter respiration contributed substantially to pre-treatment C because litter mass would have been replaced by about 95 % (3 times the turnover time). Therefore, it can be assumed that most of the fraction of soil-respired CO_2 (i.e. in total R_S) derived from pre-treatment C pools (i.e. C assimilated before 1996; Eq. 4) originated from SOM decomposition (including some root litter decomposition). Therefore, the contributions of $C_{\text{pre-tr}}$ (oxidation of soil pools older than 8 yr) and $C_{\text{post-tr}}$ were applied to the rate of SOM decomposition.

2.8 Canopy air temperature and soil temperature and moisture

Continuous temperature measurements were taken at lower canopy air and at 10 cm soil depth in each FACE plot, using Siemens Type M 841/S1 thermistors at 30 min interval averages. Continuous soil moisture measurements were taken with a Campbell Scientific Model CS 615 probes (Logan, Utah, USA) consisting of two 30 cm long metal rods, over which each moisture measurement is integrated. Soil temperature and moisture measurements were taken every 5 or 30 s, averaged over 30 min intervals and automatically logged with Campbell 21X or 23X data loggers.

The rate of total R_S and each R_S component was plotted against soil temperature and soil moisture at each measurement date and time (regressions not shown). Litter respiration was also plotted against lower canopy air temperature. The relationship between soil temperature and R_S and its components was determined by fitting a second-order exponential growth function to the data, according to the equation $f = ae^{bx}$ (Lloyd and Taylor, 1994). The relationship between R_S and each R_S component and soil moisture was determined by fitting linear functions to the data, according to the equation $f = y_0 + ax$ (Orchard and Cook, 1983).

2.9 Statistical and error sensitivity analyses

Temporal variability in R_S and R_S components was examined with mixed-effects and random-effects regression analyses (Proc Mixed, SAS v. 9.1, Cary, NC). Unlike regular regressions, random-effects regression does not assume each measurement is independent, but assumes data are dependent on clusters, here FACE plots (Hedeker et al., 1994). This method also allows for analyses of unbalanced data (i.e. different observations at different clusters or time series). With

balanced datasets, this method is analogous to nested analyses of variance of mixed-model regressions. Rates of R_S in 2003 and 2004 were fitted to a regression model with CO_2 treatment, time of day, month, and year as covariates, and interactions of CO_2 treatment with time of day and year. For 2004, regression models with effects for month, time of day (day or night), month by time of day interaction, and a random effect for plot were fitted to root versus non root R_S and to R_R , R_{SOM} , $R_{\text{pre-tr}}$, $R_{\text{post-tr}}$, and R_L rates.

A sensitivity analyses was made to estimate the error propagation of the calculated R_R , R_{SOM} and R_L components of R_S to endmember determinations. Rapid variations in the isotopic composition of respired CO_2 may induce an error in the mixing models used here. For this sensitivity analyses we applied a $\pm 1.5\text{‰}$ to the root, SOM and litter respired CO_2 determinations from the incubation chambers. The isotopic composition of, particularly root respiration, can vary rapidly depending on, for instance, photosynthetic conditions. The sensitivity analyses were performed to account for this variability and for potential sampling biases and propagated errors during calculations. To minimize errors originating from R_S , we averaged the resulting R_R , R_L and R_{SOM} partitioning of R_S at 12 locations within each replication unit (i.e. each FACE ring). Each per ring average was then used to obtain the treatment average ($n = 3$). We performed these analyses and approaches for each month and time of day we calculated the root litter and SOM components of R_S .

3 Results

3.1 Seasonal and interannual variability of R_S

We measured R_S for the ambient and elevated CO_2 plots for the growing seasons of 2003 and 2004. Rates of R_S differed significantly in the two years of study ($p < 0.0001$); R_S rates in 2004 were on average 16 % higher than R_S rates in 2003 (Fig. 1). Rates of R_S in both treatment and control plots showed seasonal variability in both years of study ($p < 0.0001$) with higher R_S in the middle of the growing season (Fig. 1). Soil respiration rates were not significantly stimulated by ecosystem exposure to elevated $[\text{CO}_2]$ during 2003 ($p > 0.5$), but there was a significant CO_2 treatment effect on R_S in 2004 ($p < 0.03$; Fig. 1). Rates of R_S were on average 14 % higher under elevated $[\text{CO}_2]$ in 2004 relative to ambient CO_2 conditions. The magnitude of the CO_2 treatment effect on R_S varied diurnally and seasonally in the two years of measurement (Fig. 1). Daytime R_S rates under elevated $[\text{CO}_2]$ in 2003 were between 1 % (in August, $p > 0.8$) and 20 % (in September, $p < 0.1$) higher than daytime R_S rates under ambient $[\text{CO}_2]$. Nighttime R_S rates in 2003 were between 5 % lower (in August, $p > 0.5$) and 10 % higher (in September, $p > 0.5$) than nighttime rates under ambient $[\text{CO}_2]$. In 2004, the enhancement of daytime R_S rates in the CO_2 treatment plots was between 9 % (in August,

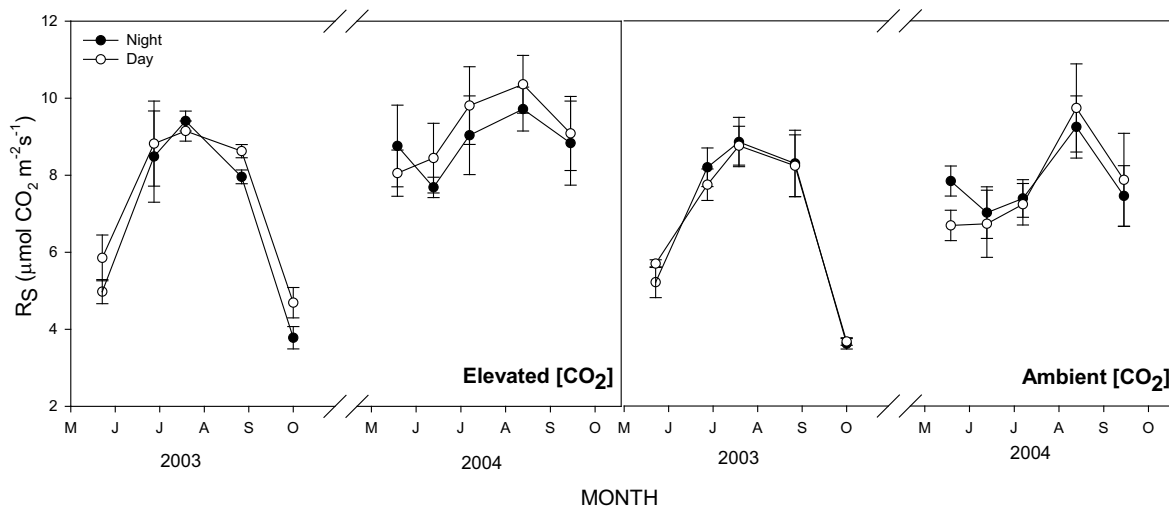


Fig. 1. The rates of soil respiration at FACTS-1 under ambient and elevated $[\text{CO}_2]$ during the growing seasons of 2003 and 2004 and measured at night (filled symbols) and daytime (open symbols). Values are means \pm standard error ($n = 3$).

$p < 0.1$) and 21 % (in July, $p < 0.002$). Nighttime R_S rates under elevated $[\text{CO}_2]$ in 2004 were between 5 % (in August, $p > 0.4$) and 17 % (in July, $p < 0.02$) higher than R_S rates in the control plots.

3.2 Nighttime and daytime R_S

Daytime and nighttime R_S rates were not significantly different during the two years of measurements ($p > 0.2$) under either ambient or elevated $[\text{CO}_2]$, with the exception of a significant CO_2 treatment \times time interaction in 2003 ($p < 0.04$; Fig. 1). In 2003, daytime R_S rates were on average 6 % higher than nighttime rates under elevated $[\text{CO}_2]$, although at ambient $[\text{CO}_2]$ conditions seasonal daytime and nighttime R_S rates differed by less than 1 %. In 2004 at elevated $[\text{CO}_2]$, daytime R_S rates were 3 % higher than nighttime rates ($p > 0.3$). At ambient $[\text{CO}_2]$ daytime rates were 3 % lower than nighttime rates under ambient $[\text{CO}_2]$ ($p > 0.2$). Daytime R_S rates were 9 % higher in 2003 and 17 % higher in 2004 under elevated $[\text{CO}_2]$ relative to ambient $[\text{CO}_2]$ ($p < 0.1$; Fig. 1). Nighttime rates of R_S were 2 % and 12 % greater under elevated $[\text{CO}_2]$ in 2003 and 2004, respectively ($p > 0.1$; Fig. 1).

3.3 Soil respiration components under elevated $[\text{CO}_2]$

The continuous whole ecosystem C-isotope label in forest plots exposed to elevated $[\text{CO}_2]$ (beginning September 1996) offered the opportunity to partitioning soil-respired CO_2 into several source components at the 7th and 8th growing seasons after high CO_2 exposure. This isotope partitioning is not possible at ambient conditions and therefore the following analyses are restricted to the treatment plots only. Soil respiration was partitioned into two C-age components in 2003 and 2004. In 2004, R_S was further partitioned into 4 source components.

3.3.1 Post-treatment C in R_S in 2003 and 2004

Pre-treatment (C fixed by the ecosystem prior to September 1996) and post-treatment C (C fixed by the ecosystem after September 1996) partitioning of respired CO_2 by soils was done during the growing seasons of 2003 (7th growing season of isotope label exposure) and 2004 (8th growing season of isotope labeling) as in Taneva et al. (2006). Post-treatment C flux was the largest flux component in R_S and showed no significant diurnal variation (Fig. 2). Post-treatment C flux had a strong seasonal pattern during the growing season of 2003 (Fig. 2a) where $R_{\text{post-tr}}$ was higher in the June–August period than in previous and posterior months. This seasonal variation was less pronounced during the growing season of 2004 where rates of $R_{\text{post-tr}}$ were higher than in 2003 at the beginning and the end of the growing season.

In 2003, the contribution of post-treatment C (from autotrophic and heterotrophic sources) to daytime R_S ranged from 58.6 ± 8.3 % in May to 87.5 ± 5.1 % in July (Fig. 2). At night, the contribution of post-treatment C to R_S ranged from 56.2 ± 12.9 % in June to 84.0 ± 3.8 % in August. In 2004, the daytime flux and contribution of post-treatment C to R_S was higher than in 2003 ($p < 0.04$) and ranged from 82.5 ± 9.1 % in May to 89.9 ± 3.5 % in July (Fig. 2). The contribution of post-treatment C to nighttime R_S was less than during the day and ranged from 76.6 ± 9.1 % in August to 84.8 ± 6.3 % in May (Fig. 2).

3.3.2 Root/rhizosphere respiration (R_R) in 2004

In 2004, R_R had a seasonal average of 2.81 ± 0.50 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ during the day and 3.04 ± 0.66 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ at night (Fig. 3; Table 2). Overall daytime and nighttime R_R rates were not significantly different ($p > 0.3$), despite

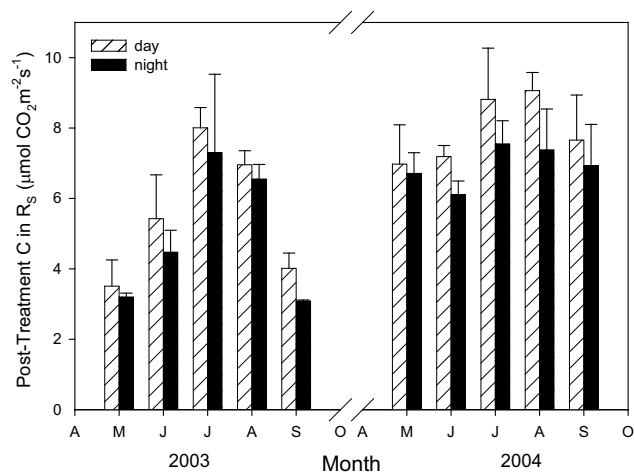


Fig. 2. The amount of post-treatment belowground C (C fixed after September 1996) to daytime (open bars) and nighttime (closed bars) soil respiration rates during the growing seasons of 2003 and 2004 at FACTS-1. Values are means ($n = 3$) \pm standard error.

significant differences in diel rates in August. Overall, R_R rates were 8% lower during the day than at night (Table 2). Significantly higher rates of R_R were observed in the middle of the season (July and August) relative to rates early or late in the season (June and September), both during the day (36%; $p < 0.001$) and at night (39%; $p < 0.0001$). These differences were mostly due to much lower daytime R_R rates in September and much higher nighttime rates in August, relative to the rest of the season (Table 2). Daytime and nighttime R_R rates in the middle of the season were also significantly different from those early or late in the season ($p < 0.0001$). The average contribution of R_R to total R_S was $29.7 \pm 5.3\%$ during the day, ranging from $14.1 \pm 4.4\%$ in September to $36.8 \pm 4.1\%$ in June (Table 2). At night, the average contribution of R_R to total R_S was $33.7 \pm 5.9\%$, ranging from $26.6 \pm 4.5\%$ in September to $51.4 \pm 3.5\%$ in August (Table 2).

3.3.3 Litter decomposition (R_L)

In 2004, R_L had a seasonal average of $2.56 \pm 0.99 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ during the day and $1.16 \pm 0.57 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ at night (Fig. 3; Table 2). Overall, daytime R_L rates were significantly different from nighttime rates ($p < 0.0001$), despite non-significant differences in diel rates in June and July (Fig. 3). On average, daytime R_L rates were 55% higher than nighttime R_L rates. Neither daytime nor nighttime rates of R_L showed seasonal variability ($p > 0.2$) in 2004.

The average contribution of R_L to total R_S was $25.7 \pm 10.5\%$ during the day, ranging from 0% in June to $51.4 \pm 6.7\%$ in September (Table 2). At night, the average contribution of R_L to total R_S was $12.4 \pm 6.2\%$, ranging from 0% in June to $24.3 \pm 6.2\%$ in July (Table 2).

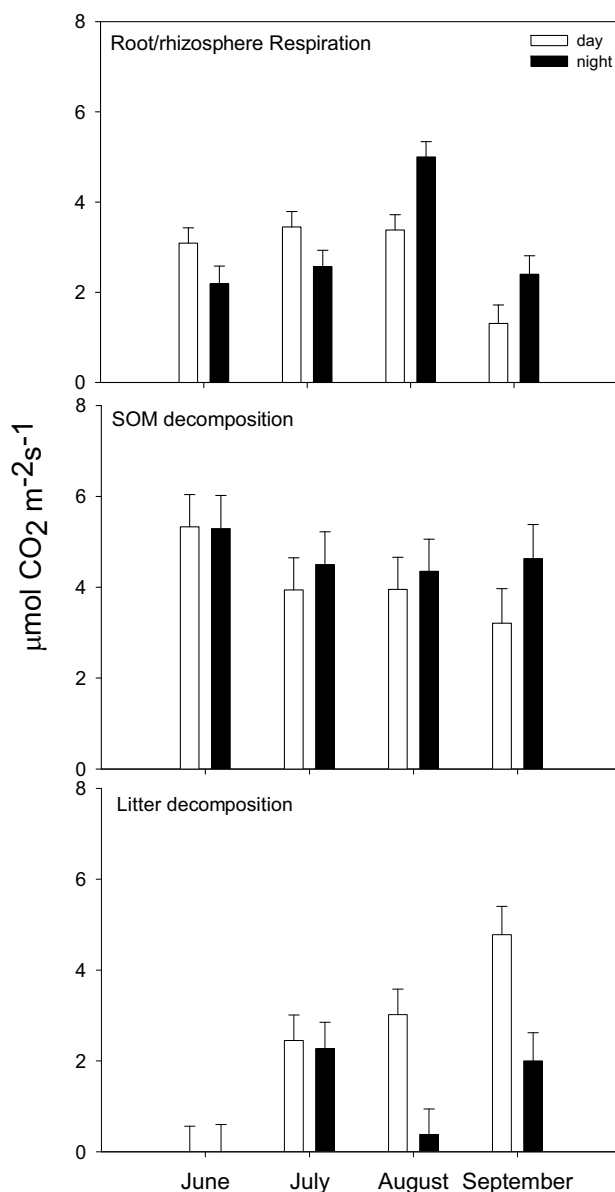


Fig. 3. The rates of the root, soil organic matter, and litter respiration components of soil respiration during the day (open bars) and at night (closed bars) in 2004 at FACTS-1. Values are means ($n = 3$) \pm standard error.

3.3.4 SOM decomposition (R_{SOM})

In 2004, R_{SOM} had a seasonal average of $4.11 \pm 0.44 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ during the day and $4.69 \pm 0.21 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ at night (Fig. 3; Table 2). There were significant differences in daytime and nighttime R_{SOM} rates ($p < 0.02$), mostly because of significant differences between daytime and nighttime R_{SOM} rates in September ($p < 0.01$; Fig. 3). Overall, daytime R_{SOM} rates were 14% lower than nighttime R_{SOM} rates. Significantly higher rates of R_{SOM} were observed early in the season (June and July)

Table 2. The CO₂ efflux rates and the relative contribution of root (R_R), soil organic matter (R_{SOM}), litter (R_L) and pre-treatment C (fixed by the ecosystem up to the year 1996; $R_{Pre-tr C}$) components of soil respiration during the day and at night in treatment plots at FACTS-1 over the growing season of 2004. Respiration rates are expressed in $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ and show average values ($n = 3$) \pm standard error.

FLUX:	R_R	% R_R	R_{SOM}	% R_{SOM}	R_L	% R_L	$R_{Pre-tr C}$	% $R_{Pre-tr C}$
DAY								
June	3.09 \pm 0.34	36.8 \pm 4.1	5.33 \pm 0.71	63.5 \pm 8.4	0.0 \pm 0.6	0.0 \pm 6.7	1.24 \pm 0.47	14.8 \pm 5.6
July	3.45 \pm 0.34	35.1 \pm 3.5	3.94 \pm 0.71	40.1 \pm 7.2	2.45 \pm 0.56	24.8 \pm 5.7	0.95 \pm 0.47	9.7 \pm 4.7
August	3.38 \pm 0.34	32.6 \pm 3.3	3.95 \pm 0.71	38.2 \pm 6.8	3.02 \pm 0.56	29.2 \pm 5.4	1.21 \pm 0.47	11.7 \pm 4.5
September	1.31 \pm 0.41	14.1 \pm 4.4	3.21 \pm 0.76	34.5 \pm 8.0	4.78 \pm 0.62	51.4 \pm 6.7	1.72 \pm 0.48	18.5 \pm 5.1
NIGHT								
June	2.19 \pm 0.39	29.3 \pm 5.2	5.29 \pm 0.73	70.8 \pm 9.7	0.0 \pm 0.6	0.0 \pm 8.0	1.11 \pm 0.47	14.9 \pm 6.3
July	2.57 \pm 0.36	27.5 \pm 3.9	4.50 \pm 0.72	48.2 \pm 7.7	2.27 \pm 0.58	24.3 \pm 6.2	1.70 \pm 0.47	18.2 \pm 5.0
August	5.00 \pm 0.34	51.4 \pm 3.5	4.35 \pm 0.71	44.7 \pm 7.3	0.38 \pm 0.56	3.9 \pm 5.8	2.17 \pm 0.47	22.2 \pm 4.8
September	2.40 \pm 0.41	26.6 \pm 4.5	4.63 \pm 0.75	51.2 \pm 8.2	2.00 \pm 0.62	22.1 \pm 6.9	2.24 \pm 0.48	24.8 \pm 5.3

relative to later in the season (August and September) during the day (23%; $p < 0.003$), mostly because of high R_{SOM} rates in June (Fig. 3; Table 2); there were no significant differences between early- and late-season nighttime R_{SOM} rates ($p > 0.2$).

The average relative contribution of R_{SOM} to total R_S was $44.1 \pm 6.6\%$ during the day, ranging from $34.5 \pm 8.0\%$ in September to $63.5 \pm 8.4\%$ in June (Table 2). At night, the average contribution of R_{SOM} to total R_S was $53.7 \pm 5.8\%$, ranging from $44.7 \pm 7.3\%$ in August to $70.8 \pm 9.7\%$ in June (Table 2).

3.4 Partitioning of R_H ($R_{SOM} + R_L$) into pre- and post-treatment components

It is unlikely that C fixed prior to 1996 will contribute to autotrophic respiration during the growing season of 2004. Therefore, all pretreatment C is likely to originate from R_H ($R_{SOM} + R_L$) during decomposition. The mean residence time of forest floor C is 2.5 yr (Lichter et al., 2008) and by 2004 the forest floor pool would have been replaced by about 95%. Accordingly pretreatment C could contribute to no more than 20% of total R_L rates, but pre-treatment C from R_L contributed by 5% of total pre-treatment C efflux in R_S . Because most of the pre-treatment C in soil CO₂ efflux originated from R_{SOM} , R_{SOM} was further distinguished between two age pools at the elevated [CO₂] plots during 2004: pre-treatment C consisting of C fixed by the ecosystem prior to September 1996 (>8 yr old), and post-treatment C, assimilated after fumigation began (<8 yr old). In 2004, the seasonal average rate of post-treatment SOM decomposition was $2.83 \pm 0.53 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ during the day and $2.89 \pm 0.45 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ at night (Fig. 4). The seasonal average rate of pre-treatment SOM decomposition was 1.28 ± 0.16 and $1.81 \pm 0.26 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ for day and night, respectively (Fig. 4).

Overall rates of post-treatment SOM-C decomposition did not significantly differ between day and night ($p > 0.7$; Fig. 4). Higher rates were seen earlier in the season (June and July), both during the day (40%, $p < 0.0001$) and at night (34%, $p < 0.001$). Unlike post-treatment SOM decomposition, the rates of pre-treatment SOM decomposition differed significantly between day and night ($p < 0.0001$), despite non-significant differences in June ($p > 0.4$). Nighttime rates of pre-treatment R_{SOM} were about 29% higher than its daytime rates (Table 2). Seasonal variability in the decomposition of pre-treatment SOM was also significant with rates earlier in the season (June and July) lower than rates later in the season (August and September), both during the day (25%, $p < 0.005$) and at night (36%, $p < 0.0001$).

The average contribution of post-treatment SOM decomposition to total R_S was $30.4 \pm 6.8\%$ during the day, ranging from $16.2 \pm 5.4\%$ in September to $48.6 \pm 5.6\%$ in June (Table 2). The average relative contribution of pre-treatment SOM decomposition to total R_S was $13.7 \pm 1.9\%$ during the day, ranging from $9.7 \pm 4.7\%$ in July to $18.5 \pm 5.1\%$ in September (Table 2). At night, the average contribution of post-treatment R_{SOM} to total R_S was $33.7 \pm 7.5\%$, ranging from $22.5 \pm 4.8\%$ in August to $55.9 \pm 6.5\%$ in June (Table 2). At night, the average contribution of pre-treatment SOM decomposition to total R_S was $20.0 \pm 2.2\%$, ranging from $14.9 \pm 6.3\%$ in June to $24.8 \pm 5.3\%$ in September (Table 2).

We also tested whether the increases seen in R_S during the summer were due to R_R or R_H (i.e. $R_{SOM} + R_L$). Both daytime and nighttime R_S rates increased during the summer ($t = 4.50$; $p < 0.0001$) compared to late spring and early fall (see the Supplement for details). Daytime and nighttime rates of R_R also increased significantly during the July and August periods compared to June and September ($t = 3.11$; $p = 0.0022$), whereas R_H did not statistically increased

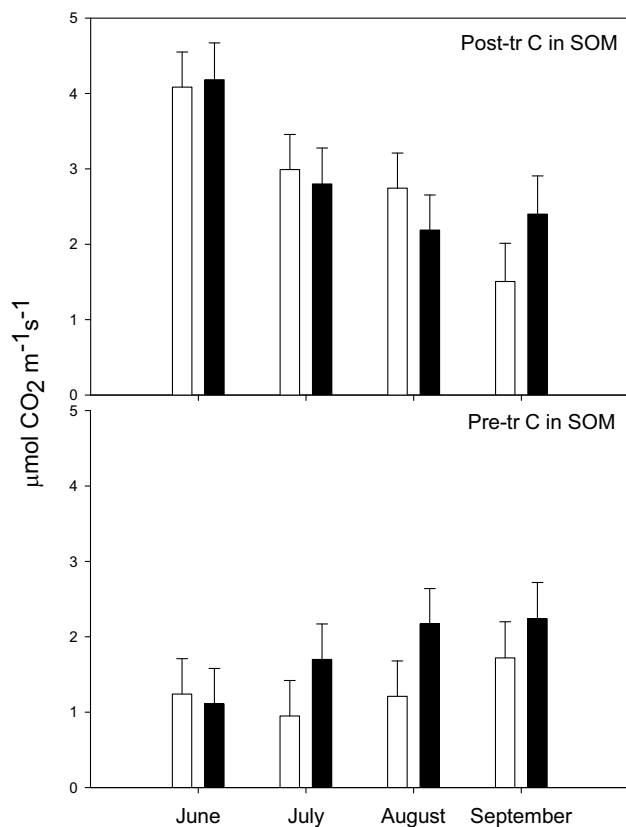


Fig. 4. The contribution of post- and pre-treatment soil organic carbon to daytime and nighttime soil respiration at FACTS-1 during the growing season of 2004. Values are means ($n = 3$) \pm standard error.

respiration rates during this period ($t = 0.74$, $p = 0.45$). Therefore, seasonal variation in R_R contributed to variations in R_S .

3.5 Temperature and moisture relationships with components of R_S

There were no significant differences in daytime and nighttime soil temperature or moisture for the measurement periods in this study (t -test, $p > 0.05$). Seasonal patterns of total CO_2 efflux were influenced by temperature ($R^2 = 0.32$) and moisture ($R^2 = 0.28$), particularly at night where combined soil temperature and moisture could explain 66% of seasonal nighttime variations in R_S ($p < 0.05$). However, at diurnal time scales these interactions were not significant, mostly due to lack of soil temperature changes between day and night (except for June 2004 where night respiration was slightly lower than daytime respiration).

In 2004, the components of soil respiration also showed little sensitivity to diurnal changes in soil temperature, as some components of R_S increased, decreased or remained unchanged diurnally. Again there was little diurnal change

in temperature between day and night, illustrating biotic interactions in diel variation of some R_S components. Moisture variations were relatively modest both diurnally and seasonally and volumetric water content remained above 0.2 (Table 3), and therefore regressions were not significant with the flux variables.

At seasonal time scales, both daytime ($R^2 = 0.78$) and nighttime ($R^2 = 0.40$) R_R were correlated with soil temperature (see the Supplement). Flux variations in the other R_S components were insensitive to temperature and moisture. Litter respiration may be more sensitive to air temperature than soil temperature. Air canopy temperature had about 6°C variation between day and night during the summer months. Litter respiration was not significantly correlated to air temperature at diurnal time scales as R_L did not exhibit a diurnal pattern. Seasonally, daytime R_L was slightly correlated with lower canopy air temperature ($R^2 = 0.26$) whereas nighttime R_L was poorly correlated with air temperature ($R^2 = 0.15$).

3.6 Sensitivity analyses for isotope and error accumulation

Error accumulation was initially minimized by applying the mixing models at the individual collar level (12 per ring). With this approach the within ring standard errors were analyzed and they were between 2.7% and 6.9% of the value of the mean depending on the R_S component, time of measurement and ring. Therefore the within ring variation in rates was similar or smaller than variation seen across rings ($n = 3$). Also the compound estimates of R_S using the isotope collar method were within 5% of R_S measurements made by other methods (e.g. Bernhardt et al., 2006; Taneva et al., 2006).

Errors can be introduced in the calculation of pre- and post-treatment C if isotopic variations at ambient rings if changes in the isotopic composition of R_S are caused by changes in the relative contribution between roots and heterotrophs. This potential error was probably small because there were small variations in the isotopic composition of root- and SOM- respired CO_2 over time (Table 1). An unlikely more than a 60% shift in the relative contribution of R_R and R_H is needed to change the isotopic composition of soil-respired CO_2 by 1‰. A sensitivity analysis considering this unlikely large abrupt change in the R_R and R_H contributions to R_S at ambient shows that it will induce a less than 6% error in the relative separation of pre- and post-treatment C at the elevated plots.

In addition, errors can be introduced with rapid variation in the isotopic composition of respired- CO_2 from roots, root-free soil or forest floor litter. It has been documented that the isotopic composition of respired CO_2 can shift relatively rapid with respect to the isotopic composition of metabolized substrate, gas diffusion, or other factors (see Vargas et al., 2011). In this experiment, the isotopic composition of

Table 3. Soil temperature at 10 cm (°C), volumetric soil water content at 10 cm and lower canopy air temperature (°C) in the treatment plots at FACTS-1. Reported values are means and standard error ($n = 3$) of the days experiments were made.

Month	Soil Temperature (°C)		Soil Moisture (% vol)		Lower Canopy Air Temperature (°C)	
	Day	Night	Day	Night	Day	Night
June	20.2 ± 0.3	20.3 ± 0.3	0.21 ± 0.01	0.21 ± 0.01	25.0 ± 0.4	18.1 ± 0.6
July	21.3 ± 0.2	21.4 ± 0.2	0.20 ± 0.02	0.20 ± 0.02	25.9 ± 0.4	19.5 ± 0.2
August	21.2 ± 0.1	21.3 ± 0.1	0.28 ± 0.03	0.27 ± 0.03	24.8 ± 0.2	17.8 ± 0.4
September	19.1 ± 0.6	19.2 ± 0.4	0.29 ± 0.00	0.29 ± 0.00	21.5 ± 0.2	17.3 ± 0.4

respired- CO_2 from each belowground source was measured rather than using the isotopic composition of a given bulk tissue or pool (Table 2). The use of measured $\delta^{13}\text{C}$ of respired CO_2 shall integrate the isotopic variations caused by intrinsic or environmental conditions for each pool and every time the R_S component partitioning was made (e.g. Hymus et al., 2005; Moore et al., 2008). Extrapolation of the measured $\delta^{13}\text{C}$ of respired CO_2 values over time (less than 24 h in this study) may induce errors if rapid isotopic variation of respired- CO_2 occurs. To account for this potential variation, we performed a sensitivity analysis for each month and time of day for which the component partitioning of R_S was done. We calculated the effect of an error of 1.5 ‰ shift in the measured versus real isotopic composition of respired CO_2 for any given endmember at any given time and analyzed the resulting component partitioning of R_S . The sensitivity analyses revealed that for every 1 ‰ change in the isotopic composition of respired- CO_2 from either roots, root-free soil or litter, the R_R component of R_S varied up to 15 %, whereas the variation in the R_{SOM} or R_L contribution to R_S changed by less than 6 % on average.

4 Discussion

In this study, we document that variations in individual components of R_S do not always lead to measurable variations in overall R_S efflux rates. Conversely, changes in R_S may not be always attributed to a one single component. We also report that diel differences in rates of R_S components are not easily explained by passive temperature and moisture controls, and that biotic controls on R_H are also important in determining rates of SOM oxidation. This is particularly evidenced by the diurnal pattern of oxidation of old C ($R_{\text{pre-tr}}$). While this is not the first study to separate R_S into more than two components (Sulzman et al., 2005; Cisneros-Dozal et al., 2006; Subke et al., 2011; Vargas et al., 2011), to our knowledge, this is among the first reports of the diel and seasonal changes in the contribution of four R_S components to growing season efflux rates under field conditions. Our results suggest that fine controls on individual components contributing to soil CO_2 efflux could result in different responses to similar biotic and abiotic variables. Understanding the sources of

soil CO_2 efflux and its dependent biotic and abiotic controls are important in elucidating the environmental effects on R_S rates at different time scales.

By taking advantage of the ecosystem ^{13}C tracer we were able to examine if and how the temporal dynamics of R_S components translate into temporal variability of total R_S . Due to the lack of a ^{13}C tracer in the control plots at the Duke Forest FACE site, we were only able to study R_S components under ecosystem exposure to elevated $[\text{CO}_2]$ and a comparison of the contributions of different R_S components under ambient and elevated CO_2 conditions was not possible. Therefore, the interpretation of results is within the constraints of the sensitivity analyses of using a single isotope, and influenced by the effects of elevated CO_2 in below ground forest dynamics.

4.1 Soil respiration and its autotrophic and heterotrophic components

Several studies have reported increased R_S rates under elevated $[\text{CO}_2]$ (King et al., 2004; Bernhardt et al., 2006; Taneva et al., 2006) and our results are in agreement with these reports for the growing season of 2004 but not for 2003 (Fig. 1). There were little cumulative climatic differences between the growing seasons of 2003 and 2004, but the growing season of 2002 was slightly dryer than normal. In 2002, both net primary productivity and belowground C allocation of the forest exposed to high CO_2 reached the lowest values since 1996 (Finzi et al., 2006), which may have caused a legacy effect on belowground processes during the following year (decreased storage C, lower root production, etc). During both growing seasons, R_S showed a strong seasonal pattern but not a diurnal pattern (Fig. 1). The presence or absence of diurnal variability of total R_S rate could not be attributed to variability in the rate of any single R_S component (Table 3), although the seasonal variability seemed to be partly driven by changes in R_R .

The contribution of R_R to total R_S ranged from 14 to 37 % during the day in this forest. This range is on the lower end of the annual range of 20–84 % reported for temperate coniferous forests (reviewed by Subke et al., 2006) and lower than previous annual estimates at the site using midday rates of R_S (Andrews et al., 1999; Hamilton et al., 2002). The proportion

of R_R in R_S from this study is consistent with the relatively low levels of root productivity and turnover as compared to other temperate forests (Matamala and Schlesinger, 2000; Matamala et al., 2003). Subke et al. (2006) reported a relative increase in the contribution of R_R to R_S as the rate of R_S increases, as is the case of this study (Table 3), suggesting that R_R may influence R_S during the growing season (see the Supplement, Vargas et al., 2011).

Although we found R_R to be temperature sensitive over seasonal time scales, R_S was poorly correlated to changes in soil temperature or moisture at both diurnal and seasonal time scales. There is a growing body of evidence that R_S shows a diurnal hysteretic response in many ecosystems (Vargas et al., 2011), suggesting biotic interactions may mask abiotic controls on R_S . Hysteresis in R_S has now been reported in a variety of ecosystems (Gaumont-Guay et al., 2006; Barron-Gafford et al., 2010; Philips et al., 2011), and may be caused by changes in photosynthesis, use of C and N reserves, hydraulic lift, or phenology (Högberg et al., 2001; Barron-Gafford et al., 2010; Vargas et al., 2011). These factors could affect rates of R_R which will translate into rates of R_S as a function of the relative contribution of R_R to R_S . It is less known how biotic interactions affect diurnal and seasonal patterns of R_H .

Heterotrophic respiration was the dominant component of growing season R_S in this forest, constituting 63 to 86% of daytime R_S rates (Table 2), within the reported range of 16–80% for the contribution of R_H to R_S in temperate coniferous forests (Subke et al., 2006). Heterotrophic respiration was also fueled by a substantial contribution of post-treatment C (Table 2, Fig. 2). Although it is widely recognized that R_H can result from a number of soil C pools, R_H is usually treated as a single R_S component. In this study, we further partitioned R_H into litter decomposition (R_L) and SOM decomposition (R_{SOM} , Table 2). The proportion of R_L in R_S ranged from 0 to 51% of total R_S , which is consistent with a range of 1 to 42% of R_S shown during the growing season of a temperate deciduous forest (Cisneroz-Dozal et al., 2006). This variability in the proportion of R_L in R_S may be due to changes in forest floor moisture content (Hanson et al., 2003; Goulden et al., 2004; Lee et al., 2004). However we found only a modest relationship between R_L rates and soil moisture and air temperature, suggesting other possible controls on R_L in this forest (see Malcom et al., 2009). One such control on R_L could be litter mass (unfortunately forest floor mass variation was not measured). Litter decomposition rates are expressed on per ground area basis and changes in litter mass can affect the per ground area litter flux. Although R_L on a per mass basis can respond to moisture changes as reported earlier (e.g. Hanson et al., 2003) changes in litter mass over the course of the growing season can result in different seasonal rates of R_L , masking the temperature and moisture sensitivities of litter decomposition in this forest.

4.2 Temporal variability in total R_S and R_S components

While greater rates of R_R in July and August were correlated with increased R_S , the contribution of R_H (R_{SOM} and R_L) to R_S was not constant during the growing season (Table 2), indicating that seasonal variability in R_R alone cannot explain the seasonal variability in R_S . Notably, when R_S rates were at their highest, the rates of both R_R and R_{SOM} increased. These results indicate that increases in overall R_S rates are not always solely driven by root and rhizosphere activity, as seen in other studies (e.g. Högberg et al., 2001). Different soil C pools may interact to produce observed rates of R_S and measurements of soil CO_2 efflux alone cannot account for the variability of and interactions between R_S components (Trueman and Gonzalez-Meler, 2005).

Although daytime R_S did not differ significantly from nighttime rates, there were significant diel changes in individual R_S components (Figs. 1 and 3). All three components of R_S (R_R , R_{SOM} , and R_L) had significantly different rates between day and night, although the magnitude and direction of the rate difference varied throughout the growing season. For instance, the contribution of R_R to total R_S was greater at night later in the season than during the day (Table 2). The observed diel differences in R_R are likely the result of the diurnal variability in the allocation of photosynthetic C to roots (Trueman and Gonzalez-Meler, 2005; Tang et al., 2005) and not necessarily to just changes in soil temperature and moisture; changes in daytime and nighttime values of soil temperature and moisture were small to detect their effects on diurnal variations in rates (Table 3). Because the diel differences in R_R were not always significant in this study, the time lag between photosynthesis and R_R may not be constant during the growing season of this forest (Tang et al., 2005; Vargas et al., 2011; but see Stoy et al., 2007).

Indirect evidence has shown that R_R may be a possible driver of R_S variability (Högberg et al., 2001; Bond-Lamberty et al., 2004; Trueman and Gonzalez-Meler, 2005; Subke et al., 2006) including this study site (Palmroth et al., 2006). In this study, variability in R_R alone was insufficient in explaining the seasonal and diel variability of R_S because temporal changes in other R_S components could compensate for changes in R_R . The average relative contribution of R_{SOM} to R_S was greater at night than during the day (Table 2). Since total rate of R_S did not differ between day and night, nighttime decreases in R_R , perhaps due to the absence of photosynthesis, were compensated for by increases in nighttime rates of R_{SOM} , increasing its proportion in R_S . However, lower R_R did not always translate to higher R_{SOM} during the nighttime (Fig. 3 and Table 2). Whether these variations were independent or the result of more complex interactions needs further study, but points to complex heterogeneity in pools contributing to R_H ($R_{SOM} + R_L$). As indicated above, the rate of R_L was the smallest of the components that contribute to the rate of total R_S (Fig. 3) and therefore unlikely to be the major player in causing R_S variability.

In contrast, R_{SOM} was the largest component contributing to R_S and it is also the more heterogeneous soil pool. Soil organic matter is often partitioned into static size and/or chemical fractions that often correspond with C average age (e.g. O'Brien et al., 2011). At the Duke FACE study site, C fixed after fumigation started has been incorporated in almost all soil fractions (Matamala et al., 2003; Lichter et al., 2008), but the measured mean residence time of these soil C pools contrast with the shorter time-scales at which pre- and post-treatment C contributed to soil CO_2 and soil respired CO_2 from these pools (Taneva et al., 2006), suggesting heterogeneous lability within a soil C pool in this forest. In 2004, pre-treatment C in R_S (fixed by the ecosystem prior to 1996) mostly originated from heterotrophic sources and represents a C pool with close to decadal and longer mean residence times. Pre-treatment C respiration (R_{pre-tr}) may contribute to up to 20% of total R_L but because R_L represented less than 20% of seasonal R_S (see above, Tables 1 and 2), 95% of pre-treatment C in R_S should originate from R_{SOM} . Therefore, R_{SOM} can be further partitioned into pre- and post-treatment C respiration.

Within R_{SOM} , the decomposition of C older than 8 yr (R_{pre-tr}) showed more pronounced and consistent diel differences than any other R_S component, with nighttime rates on average 29% higher than its daytime rates. In contrast, the decomposition of post-treatment C ($R_{post-tr}$) did not differ between day and night, suggesting that the variability in R_{SOM} appear to be due to changes in the decomposition of older C pools, rather than the decomposition of recently added SOM. The decomposition of pre-treatment older C increased over the course of the growing season. The diel sensitivity of soil C pools older than 8 years suggests that all SOM pools can rapidly respond to ecosystem exposure to environmental change through their biotic and abiotic controls (as suggested by Vargas et al., 2011).

The decomposition of older soil C pools constituted a substantial fraction of total R_S during the growing season of this forest (Table 2) and appeared to be as variable as R_R . Our results indicate that plant activity may exert a direct and/or indirect control over R_S through cascading effects on other R_S components beyond R_R . Plant activity has been previously linked to greater rates of SOM decomposition (Kuzyakov and Cheng, 2001; Kuzyakov, 2002; Subke et al., 2004; Trueman and Gonzalez-Meler, 2005) and an increasing number of studies have indicated that R_S components are not independent of each other, but have interactive effects on R_S (Trueman and Gonzalez-Meler, 2005; Vargas et al., 2011). These studies indicate that predicted increases in above- and below-ground NPP with elevated $[CO_2]$ may not necessarily translate into greater soil C storage, as increases in plant activity may simultaneously increase the decomposition of recent and older C in forests (Hoosbeek et al., 2004; Subke et al., 2004; Sulzman et al., 2005; Trueman and Gonzalez-Meler, 2005). Despite the importance of potential priming of old SOM decomposition by enhanced plant activity with chang-

ing environmental conditions, mechanisms of this priming remain poorly understood (Trueman and Gonzalez-Meler, 2005; Kuzyakov and Gravrichkova, 2010).

The presence of unpredictable diel patterns in the rates of R_S components, with no changes in soil temperature or moisture, suggests that primary and secondary responses of decomposers to changes in soil conditions exist (Trueman and Gonzalez-Meler, 2005; Vargas et al., 2011) including the physiological thermal acclimation of decomposers (Bradford et al., 2008). Therefore, extrapolation of daytime measurements of R_S to monthly or annual scales or application of growing season Q_{10} values to annual R_S may introduce a bias in long-term ecosystem C budgets. In our study, decomposition of SOM, particularly pre-treatment SOM (R_{pre-tr}), was the only R_S component that exhibited consistently higher contribution to R_S at night, which increased towards the end of the growing season, despite no significant differences in intrinsic decomposition kinetics between C_{pre-tr} and $C_{post-tr}$ at FACTS-1 (Taneva and Gonzalez-Meler, 2008) or other studies (Trueman et al., 2009). These results suggest that the oxidation of older SOM may be affected by short-term environmental or biotic controls that may result from interactions between plant and decomposer activity. It is possible that root activity during the daytime interacts with decomposition of older SOM through competition for nutrients and water or through the availability of rhizodeposits (e.g. Kuzyakov and Cheng, 2001).

In summary, the results from these experiments show that the lack of diel changes in total R_S cannot be interpreted as a sign that source components within R_S do not vary. Conversely, because the diel changes in the four components of R_S we measured were not consistent, the seasonal variation seen in R_S for this forest cannot be attributed to proportional variation within these components. Although our results are constrained by the single isotope approach and by the elevated $[CO_2]$ conditions, these results suggest that there are interactions between components of R_S at both diel and seasonal time scales. Although the nature of these interactions could not be elucidated here, they influence the temperature- and moisture-dependent functions of total R_S , as soil organisms and roots are likely to actively modulate their activity rather than passively respond to biotic and abiotic factors. Understanding these interactions and how they may elicit the decomposition of old stored soil C in response to changing environmental conditions is paramount to elucidate the effects of R_S and its components on the atmospheric concentration of greenhouse gases.

Supplementary material related to this article is available online at:
<http://www.biogeosciences.net/8/3077/2011/bg-8-3077-2011-supplement.pdf>

Acknowledgements. This research was funded by NSF to MAG-M and by a University of Illinois at Chicago Graduate Fellowship to L. T. We gratefully acknowledge the staff of Brookhaven National Laboratory and the Duke Forest for maintenance and operation of the FACTS-1 experiment. We thank A. Niederland, J. S. Pippen, and T. Sagimoto for help in the field and Leah Simoni, Sergey Oleynik, and Matt Lorz for assistance in the laboratory. We thank Jim Burns for his help with statistical analyses. Dave Moore, Rich J. Norby and A. Rocha provided constructive and insightful comments on earlier versions of this manuscript. The FACTS-1 experiment is supported by the Office of Science (BER), US Department of Energy.

Edited by: A. Knohl

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