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3	Draft genome of Halomonas strain GFAJ-1 (ATCC BAA-2256)
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18	Running title "Genome of Halomonas strain GFAJ-1"
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21 Abstract

22 Halomonas strain GFAJ-1 was reported in Science magazine to be a remarkable microbe 23 for which there was "arsenate in macromolecules that normally contain phosphate, most 24 notably nucleic acids." The draft genome of the bacterium was determined (NCBI 25 accession numbers AHBC01000001 through AHBC01000103. It appears to be a typical 26 gamma proteobacterium. 27 28 Halomonas is a diverse genus of halophilic, alkalophilic gamma proteobacteria (2). 29 Strain GFAJ-1 (13) has not been assigned a species name. The motivation for isolation of 30 strain GFAJ-1 was to find "shadow life", which is defined in Wikipedia 31 (http://en.wikipedia.org/wiki/Shadow life) as life on earth "which has no evolutionary 32 connection with life currently known to science" (perhaps cells without DNA or ribosomes). 33 There is no evidence for "shadow life". 34 When the initial report on strain GFAJ-1 was published (13), there was an immediate 35 negative reaction (e.g. 4, 7, 10, 12) to the claim of "arsenate in macromolecules that 36 normally contain phosphate, most notably nucleic acids", arguing about the inadequacy of 37 the data supporting the claim and the expected instability of arsenate di-ester bonds. No 38 progress has occurred on these matters since the initial publication. The Oremland 39 laboratory has published on novel arsenic metabolism of other species (5, 6,11). The 40 genome of strain GFAJ-1 was determined in an effort toward further understanding. The 41 genome indicates that strain is a gamma proteobacterium, the same class that includes 42 *Escherichia coli.* There is no indication in the genome of any unusual or unexpected 43 metabolism. However, the genome does not directly address the basic problem.

44	Halomonas strain GFAJ-1was grown in the medium described (13) with 1.5 mM
45	phosphate but no added vitamins or tungsten. The medium was supplemented with 0.2 g/l
46	yeast extract, 10 mM KCl, and 10 mM potassium glutamate. The strain grows rapidly, with
47	a doubling time of 2.5 hours and approximately $10^9$ cells/ml after overnight incubation at
48	29 $^{\circ}\mathrm{C}$ (data not shown). Cells in late log phase growth were harvested and lyzed by EDTA,
49	lysozyme, and detergent treatment, followed by proteinase K and RNAse digestion. DNA
50	isolation was by phenol/chloroform/isoamyl alcohol extraction and repeated
51	isopropanol/ethanol precipitation (8). DNA purity was measured as A260nm/A280nm ratio
52	and a single DNA band of size over 20 kbp was observed (data not shown) after agarose gel
53	electrophoresis. The genome was sequenced using the Illumina Hiseq 2000 sequencing
54	platform, with a random subset of 3.5 million paired-end reads (175x coverage) used for
55	assembly with MIRA V3.4rc2 into 103 contigs that were submitted to GenBank.
56	Strain GFAJ-1 was initially (13) placed in genus Halomonas based on the sequence of
57	its 16S rRNA gene; the genome sequence includes AHBC01000086.1, nte 475-2006, that is
58	identical to that in ref. 13, with a single exception in the 5' "PCR primer" that was used
59	(13). The Halomonas strain genome projects currently published are for Halomonas sp.
60	strain TD01 (3) (NCBI accession AFQW00000000.1) and for <i>H. elongata</i> strain DSM 2581
61	(9) (NCBI sequence NC_014532.1); the genome of strain GFAJ-1 appears closely related to
62	that of Chromohalobacter salexigens strain DSM 3043 (1). It is of interest to analyze
63	potential genes involved in arsenic metabolism and resistance. The predicted protein-
64	encoding genes do not include now-standard ars-gene operon of other proteobacteria,
65	including E. coli. In particular, there appears to be an absence of genes for the ArsB
66	arsenite efflux membrane protein and the ArsC arsenate reductase enzyme.

67	Nucleotide sequence accession numbers. The 3,624,896 nte in 103 contigs, 3341 CDS plus
68	+ 68 RNAs = 3409 genes draft genome of <i>Halomonas</i> strain GFAJ-1 was deposited in
69	GenBank (http://www.ncbi.nlm.nih.gov/projects/WGS/WGSprojectlist.cgi) under accession
70	numbers AHBC01000001 through AHBC01000103.
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78	REFERENCES
79	1. Arahal, D.R., M.T. Garcia, C. Vargas, D. Canovas, J.J. Nieto, and A. Ventosa. 2001.
80	Chromohalobacter salexigens sp. nov., a moderately halophilic species that includes
81	Halomonas elongata DSM 3043 and ATCC 33174. Int.J. Syst. Evol. Microbiol. 51:1457-
82	1462.
83	
84	2. Arahal, D.R., R.H.Vreeland, C.D Litchfield, M.R. Mormile, B.J.Tindall, A. Oren, V.
85	Bejar, E. Quesada, and A. Ventosa. 2007. Recommended minimal standards for describing
86	new taxa of the family Halomonadaceae. Int. J. Syst. Evol. Microbiol. 57:2436-2446.
87	
88	3. Cai, L., D. Tan, G. Aibaidula, XR. Dong, JC. Chen, WD. Tian, and GQ. Chen.

89	2011. Comparative genomics study of polyhydroxyalkanoates (PHA) and ectoine relevant		
90	genes from Halomonas sp. TD01 revealed extensive horizontal gene transfer events and co-		
91	evolutionary relationships. Microb. Cell Factories 10:88. doi:10.1186/1475-2859-10-88		
92			
93	4. Fekry, M.I., P.A. Tipton, and K.S. Gates. 2011. Kinetic consequences of replacing the		
94	internucleotide phosphorus atoms in DNA with arsenic. ACS Chemical Biology 6:127-130.		
95			
96	5. Kulp, T.R., S.E. Hoeft, M. Asao, M.T. Madigan, J.T. Hollibaugh, J.C. Fisher, J.F. Stolz,		
97	C.W. Culbertson, L.G. Miller, and R.S. Oremland. 2008. Arsenic(III) fuels anoxygenic		
98	photosynthesis in hot spring biofilms from Mono Lake, California. Science 321:967-970.		
99			
100	6. Oremland, R.S., T.R. Kulp, J.S Blum, S.E. Hoeft, S. Baesman, L.G. Miller, and		
101	J.F. Stolz. 2005. A microbial arsenic cycle in a salt-saturated, extreme environment.		
102	Science 308:1305-1308.		
103			
104	7. Rosen, B.P., A.A. Ajees, and T.R. McDermott. 2011. Life and death with arsenic.		
105	Arsenic Life: an analysis of the recent report "A bacterium that can grow by using		
106	arsenic instead of phosphorus". Bioessays 33:350-357.		
107			
108	8. Sambrook J., and D.W. Russell. 2001. Molecular Cloning: a laboratory manual.		
109	3 <sup>rd</sup> ed Cold Spring Harbor Laboratory Press, Cold Spring Harbor Laboratory NY.		
110			

111	9.	Schwibbert, K., A. Marin-Sanguino, I. Bagyan, G. Heidrich, G. Lentzen, H. Seitz,
112	]	M. Rampp, S.C. Schuster, HP. Klenk, F. Pfeiffer, D. Oesterhelt, and HJ. Kunte.
113	,	2011. A blueprint of ectoine metabolism from the genome of the industrial producer
114		Halomonas elongata DSM 2581. Environ. Microbiol. 13:1973-1994. DOI:
115		10.1111/j.1462-2920.2010.02336.x PMCID: PMC3187862.
116	10.	Silver, S., and L.T. Phung. 2011. Novel expansion of living chemistry or just a
117	\$	serious mistake? FEMS Microbiol. Lett. 315:79-80.
118		
119	11.	Stolz, J.F., P. Basu, J.M. Santini, and R.S. Oremland. 2006. Arsenic and selenium
120	j	n microbial metabolism. Annu. Rev. Microbiol60:107-130.
121		
122	12.	Tawik, D.S., and R.E. Viola. 2011. Arsenic replacing phosphate: alternative life
123	(	chemistries and ion promiscuity. Biochemistry 50:1128-1134.
124		
125	13.	Wolfe-Simon, F., J. Switzer Blum, T.R. Kulp, G.W. Gordon, E.E. Hoeft, J. Pett-
126	]	Ridge, J.F. Stolz, S.M. Webb, P.K. Weber, P.C.W. Davies, A.D. Anbar and R.S.
127		Oremland. 2011. A bacterium that can grow by using arsenic instead of phosphorus.
128	1	Science 332:1163-1166. June 3, 2011 issue, 6 months after being published on-line (2
129	]	December 2010 www.sciencexpress.org DOI: 10.1126/SCIENCE.1197258).