

Prevalence of MRSA Colonization among Neonatal and Pediatric ICU Patients in Chicago

BY

ROSIE D. LYLES

B.S., Texas Woman's University, 1999
M.D., St. Matthew's School of Medicine, 2004
M.H.A., St. Joseph's College, 2006

THESIS

Submitted as partial fulfillment of the requirements
for the degree of Master of Science in Clinical and Translational Science
in the Graduate College of the
University of Illinois at Chicago, 2015

Chicago, Illinois

Defense Committee:

Jack Zwanziger, PhD Chair and Advisor
Michael Lin, MD, MPH Medicine and Infectious Diseases
William E. Trick, MD Collaborative Research Unit

This thesis is dedicated to my mother, Ella Flowers, without whom it would never have been accomplished.

ACKNOWLEDGMENTS

I would like to thank my thesis committee--Drs. Jack Zwanziger, Michael Y. Lin, and William E. Trick--for their unwavering support and assistance. They provided guidance in all areas that helped me accomplish my research goals and enjoy myself in the process. I would also like to acknowledge Dr. Robert A. Weinstein, who is the principal investigator for the Chicago Antimicrobial Resistance and Infection Prevention Epicenter (C-PIE) for the Centers for Disease Control and Prevention; he oversaw the development of the proposal and made contributions important to the conduct of the study.

I would like to thank infection preventionists at all participating hospitals—Laura Bardowski, Cate Berends, Amanda Bonebrake, Judy Bova, Annie Braggs, Stephanie Burtun, Cari Coomer, Theras Chou, Delia DeGuzman, Onofre Donceras, Silva Garcia-Houchins, Gerry Genovese, Edward Goodwin, James Kerridge, Jean Kirk, Robin Larson, Mary Alice Lavin, Susan Lee, Jan Lepinski, Sandra Myrick, Anna O'Donnell, Maria Perez, Barbara Schmitt, Carol Schultz, Chris Silkaitia, Annie Thompson, and Kate Wickman. I would also like to thank hospital epidemiologists for their participation, including Maureen Bolon, Andrew Cha, Emily Landon, James Malow, Sunita Mohapatra, John Segreti, Stephen Weber, Sharon Welbel, and Teresa Zembower.

RDL

TABLE OF CONTENTS

<u>CHAPTER</u>	<u>PAGE</u>
I. INTRODUCTION.....	1
II. METHODS.....	3
A. Facility and patient recruitment.....	3
B. Ethical review	4
C. Culture and Data collection	4
D. Laboratory methods	4
E. Statistical analyses	5
III. RESULTS.....	6
A. Hospital-reported surveillance for MRSA.....	6
B. Point prevalence survey results.....	8
C. Molecular epidemiology and mupirocin resistance.....	10
D. Epidemiologic differences in MRSA colonization between NICU and PICU..	10
E. Appropriate contact precautions in the setting of active surveillance.....	11
F. Impact of periodic active surveillance in NICUs.....	13
G. Performance of different body sites for MRSA colonization testing.....	13
IV. DISCUSSION.....	15
CITED LITERATURE.....	18
VITA.....	20

LIST OF TABLES

<u>TABLE</u>	<u>PAGE</u>
I. PATIENT CHARACTERISTICS OF STUDY COHORT AT THE TIME OF POINT PREVALENCE SURVEY.....	6
II. PERFORMANCE CHARACTERISTICS OF INDIVIDUAL BODY SITE TESTING FOR MRSA COLONIZATION.....	14

LIST OF FIGURES

<u>FIGURE</u>	<u>PAGE</u>
1. Estimated MRSA colonization prevalence trend for NICU and PICU during the five year study period.....	9
2. The distribution of MRSA-colonized patients across ICU days of surveillance	12

LIST OF ABBREVIATIONS

CDC	Centers for Disease Control and Prevention
ICU	Intensive Care Unit
MIC	Minimum Inhibitory Concentration
MRSA	Methicillin-resistant <i>Staphylococcus aureus</i>
NICU	Neonate Intensive Care Unit
PICU	Pediatric Intensive Care Unit
PCR	Polymerase Chain Reaction
PPS	Point Prevalence Survey

SUMMARY

Background: Beginning October 2007, the MRSA Screening and Reporting Act (210 ILCS 83/) mandated active surveillance for all ICU patients in Illinois, with isolation of MRSA-colonized patients. We assessed MRSA colonization prevalence among neonatal (NICU) and pediatric (PICU) patients using a series of point prevalence surveys.

Methods: Chicago hospitals with NICU or PICU patients were recruited for 6 single-day point prevalence surveys (PPSs) approximately 6 months apart from June 2008 to July 2011. After 2011, yearly surveys were obtained in 2012 and 2013. All ICU patients were cultured for MRSA (nose and umbilicus for neonates; nose and groin for pediatric patients) using a single swab for each body site. Hospital-reported admission screen results (i.e. 210 ILCS 83/-mandated) were also obtained. Point prevalence cultures were screened for MRSA using broth enrichment, chromogenic agar, and standard confirmatory methods.

Results: All eligible hospitals (N=10) participated (10 NICUs and 6 PICUs) with 99.6% of NICU and 93% of PICU eligible patients cultured across PPSs. Hospital-reported adherence to admission screens mandated by 210 ILCS 83/ was high (99.6% for NICU and 93.3% for PICU). Overall MRSA prevalence by PPSs in NICUs was 4.2% (89/2101); PICU, 5.7% (36/632). MRSA colonization prevalence declined in NICUs (estimated yearly odds ratio [OR], 0.92, 95% confidence interval [CI] 0.77 to 1.11, $P<0.39$) but not in PICUs (OR 1.25, 95% CI 1.10 to 1.42, $P<0.001$).

Conclusion: In the time period following implementation of mandatory active surveillance, we found MRSA colonization in a proportion of NICU and PICU patients, with evidence of ongoing MRSA transmission.

I. INTRODUCTION

Methicillin-resistant *Staphylococcus aureus* (MRSA) is an important healthcare-associated pathogen among critically ill children [1]. MRSA has been well described among patients in neonatal and pediatric intensive care units (NICUs and PICUs), with many single center as well as regional outbreaks reported [2-4]. MRSA-colonized neonates and children are at risk for subsequent invasive disease, leading to significant cost and morbidity [5].

The epidemiology of MRSA, particularly with respect to colonization, is incompletely understood in the critically ill pediatric population. Few studies have surveyed for asymptomatic carriage of MRSA [6-10]. Furthermore, prior epidemiologic studies of MRSA colonization among critically ill children have been primarily single center, some during MRSA outbreak periods, limiting generalizability. Since 2000, the epidemiology of MRSA among hospitalized children has evolved in many geographic regions with the encroachment of community-associated MRSA strains.

In October 2007, Illinois became the first state in the United States to mandate active surveillance of MRSA for patients admitted to intensive care units (MRSA Screening and Reporting Act, 210 ILCS 83/). Following the start of the law, we initiated a series of regional point prevalence surveys (8 surveys over 5 years) to assess the epidemiology of MRSA colonization among NICU and PICU patients across all hospitals, community and academic, in a large metropolitan region (Chicago, IL). We evaluated whether the prevalence of MRSA

colonization would decline during the 3 years following the active surveillance mandate. We also pursued a broader goal of describing the epidemiologic characteristics of MRSA colonization in the neonatal and pediatric ICU populations, including microbiologic characterization of MRSA isolates and assessment of hospital surveillance practice.

II. METHODS

A. Facility and Patient Recruitment

In 2008, we invited acute care hospitals in the city of Chicago with NICUs or PICUs to participate in serial point prevalence surveys for MRSA colonization. Eight serial point prevalence surveys were performed over 5 years (June 21, 2008 to July 9, 2013). All NICU or PICU patients present at the time of the surveillance visit were eligible for participation. Patients who were physically unavailable at the time of the surveillance (e.g., because they were away from their room for testing or procedures) were excluded. Written informed consent was waived for this study. Parents at the bedside, as well as minors, were asked to give verbal assent using a standardized script explaining the rationale of the project.

B. Ethical Review

This project underwent ethical review at the CDC and was determined to be a non-research activity. Thus, it was not subject to a review by the CDC institutional review board. The project was also evaluated independently at each participating healthcare facility and either deemed a public health assessment or human subjects research and approved by local review boards where applicable.

C. Culture and Data Collection

On each point prevalence survey day, facilities were provided all standardized culturing and data tracking materials as well as on-site training and coordination. Local hospital staff (primarily infection preventionists, with help from ward nurses if available) as well as one investigator (RL) performed the patient-level specimen collection.

Patients were cultured for MRSA at 2 body sites (nose and umbilicus for neonates; nose and inguinal region for pediatric patients) using a single swab for each body site. A sterile dry rayon swab (BBD BBL Culture Swab, Fisher Scientific, Pittsburg, PA) was placed in one nostril and rotated 3 times such that the entire swab was moistened by the anterior nares. For neonates, a swab specimen from a 3 x 3 cm region in the umbilicus region was collected with a second sterile dry rayon swab. Similarly, for pediatric patients, a swab specimen from a 10 x 10 cm region in the inguinal region was collected with a sterile dry rayon swab. All swabs were transported to the central laboratory in liquid Stuart medium.

At the time of specimen collection, the following patient characteristics were recorded: patient age, ICU length of stay, gender, mechanical ventilation, and contact precautions status.

D. Laboratory Methods

All specimens were transported to a central laboratory and processed within 6 hours of collection. Individual nose, groin, or umbilicus swab specimens were cultured into separate tubes

of Tryptic Soy Broth with 6.5% salt (Remel, Lenexa, KS). After overnight incubation, the broth was inoculated onto chromogenic MRSA-Select agar (BioRad, Hercules, CA). After subculture, *Staphylococcus aureus* was confirmed by colonial morphology and standard biochemical techniques. Susceptibility to oxacillin was determined by the cefoxitin disk diffusion method according to Clinical and Laboratory Standards Institute (CLSI) recommendations (CLSI M7-A8). All MRSA isolates were subtyped by pulsed-field gel electrophoresis. Community-associated MRSA genotypes were defined as USA300, 400, 1000, or 1100 [11]. Mupirocin resistance (low and high level) was detected using an E-test method. High-level mupirocin resistance was defined as a minimum inhibitory concentration (MIC) ≥ 512 mcg/mL, low-level mupirocin resistance was defined as an MIC 8-64 mcg/mL, and mupirocin susceptibility was defined as MIC ≤ 4 mcg/mL [12].

E. Statistical Analyses

For bivariable comparisons, we used Fisher's exact test or Student's *t* test, as appropriate. Exact binomial methods were used to calculate 95% confidence intervals for proportions. We constructed linear regression models using the generalized estimating equations to model prevalence and incidence trends, accounting for ICU and hospital-level correlation across time. All data were analyzed using SAS software, version 9.2 (SAS Institute, Cary, NC).

III.RESULTS

In total, 10 of 10 eligible hospitals (representing all eligible 10 NICUs and 6 PICUs) voluntarily participated in the point prevalence surveys. The median NICU size was 38.5 beds, ranging from 10 to 88, and the median PICU size was 12 beds, ranging from 7 to 42.

The overall participation rate among eligible NICU patients was high across the survey period, 99.6% (1577/1584); for PICU patients, 93.3% (485/520). Patient demographics are shown in Table I.

TABLE I

PATIENT CHARACTERISTICS OF STUDY COHORT
AT THE TIME OF POINT PREVALENCE SURVEY

Covariate	NICU	PICU	P value
Length of stay, median (IQR)	18 (6 – 43)	7 (3 – 20)	<0.001
Male (%)	1141/2095 (55)	350/633 (55)	0.79
Age, median (IQR)	19 days (7 – 42)	1.8 years (0.4 - 10)	<0.001
Ventilated (%)	422/2092 (20)	182/631 (29)	<0.001
Contact isolation (%)	114/2096 (5)	266/632 (42)	<0.001

A. Hospital-reported Surveillance for MRSA

All 10 hospitals reported that they complied with the 210 ILCS 83/ legislation by performing active surveillance testing for MRSA colonization.

MRSA screening practice varied among NICUs. Of the 10 NICUs, 9 performed active surveillance for all patients at the time of admission; 1 NICU did not perform admission surveillance but instead performed weekly periodic surveillance for all patients. MRSA screening was performed using nares culture for 9 NICUs, while 1 NICU performed combined nares/axilla/groin culture using a single swab. Polymerase chain reaction (PCR)-based MRSA testing was used in 4 of 10 NICUs, with the remainder using a culture-based detection method. Screening at time points after admission (e.g., weekly, twice a month, or at ICU day 10) was used at some point during the study period for 7 of 10 NICUs, involving 55% (1162/2101) of all NICU patients surveyed.

In comparison, PICU MRSA screening practice was fairly homogeneous. Of the 6 PICUs, all performed active surveillance for MRSA at the time of admission using nasal sampling. One out of 6 PICUs utilized PCR-based MRSA testing. Screening at time points after admission was not routinely performed in any of the PICUs.

We found high rates of compliance with the state law by hospitals across the study period, with 95% of NICU patients and 94% of PICU patients in the study cohort receiving active surveillance testing for MRSA.

The overall admission prevalence of MRSA colonization, as reported by hospitals, was as follows: NICU, 1.5% (95% confidence interval [CI] 1.0 to 2.2%) vs. PICU, 6.9% (95% CI 4.0% to 7.8%). Admission prevalence did not significantly change across the 6 survey periods for any participating NICU or PICU.

B. Point Prevalence Survey Results

Of 2101 NICU patients who participated in the surveys, 89 (4.2%) were colonized with MRSA (95% CI 3.4 to 5.1%). The MRSA colonization prevalence among NICU patients remained stable during the study period (Figure 1A; estimated yearly odds ratio 0.92, 95% CI 0.77 to 1.11, $P=0.39$).

Of 632 PICU patients who participated in the surveys, 36 (5.7%) were colonized with MRSA (95% CI 4.0 to 7.8%). In contrast to the NICU trend, the MRSA colonization prevalence among PICU patients increased significantly over time (Figure 1B; estimated yearly odds ratio 1.25, 95% CI 1.10 to 1.42, $P<0.001$).

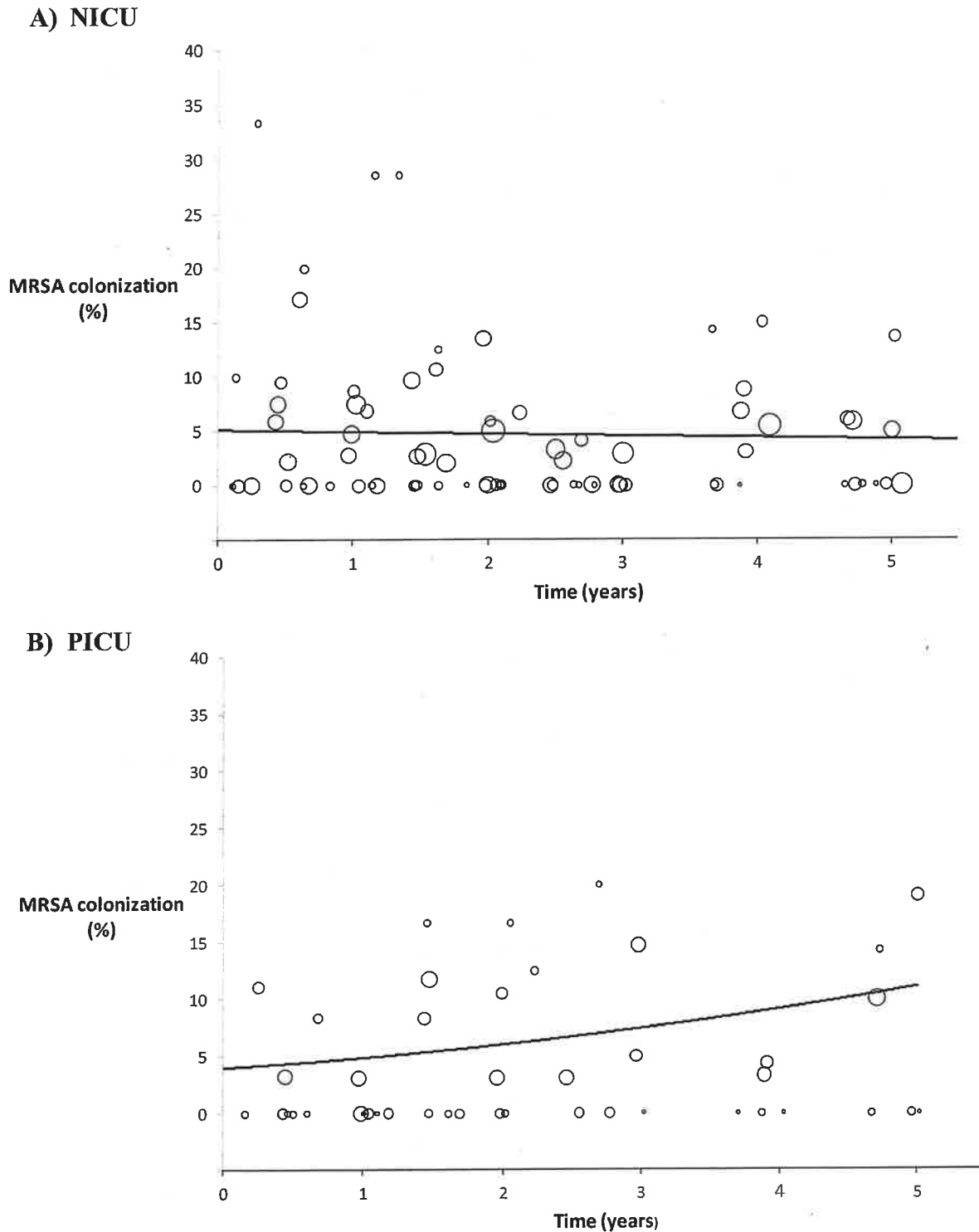


Figure 1. Estimated MRSA colonization prevalence trend for NICU and PICU during the 5 year study period. (NOTE: Each circle represents a survey point at a single ICU. Circles sizes are proportional to the relative number of patients contributing data.)

C. Molecular Epidemiology and Mupirocin Resistance

CA-MRSA genotypes represented 46% (41/89) of MRSA isolates from the NICU, and 36% (13/36) of MRSA isolates from the PICU. During the study period, the proportion of MRSA isolates represented by CA-MRSA genotypes did not change significantly for either the NICU or PICU.

In the NICU, high-level mupirocin resistance was detected in 3% (3/89) of MRSA isolates tested. Among the 36 PICU MRSA isolates tested, 8% (n=3) demonstrated high level resistance, while 3% (n=1) demonstrated low level resistance.

D. Epidemiologic Differences in MRSA Colonization Between NICU and PICU

The epidemiology of MRSA colonization differed in terms of ICU day distribution between NICU and PICU settings (Figure 2). In the NICU, MRSA-colonized neonates were detected only on ICU day ≥ 8 ; of the 192 neonates (9% of total sample) surveyed within the first 2 ICU days, none were MRSA-colonized. In contrast, MRSA-colonized PICU patients were found throughout the entire range of ICU days. The median ICU day for MRSA-colonized NICU patients was 29, versus 9 for PICU patients ($P < 0.001$).

E. Appropriate Contact Precautions in the Setting of Active Surveillance

Among MRSA-colonized patients identified through the point prevalence surveys, 44% (39/89) in the NICU and 67% (24/36) in the PICU ($P=0.03$ for difference) were in contact isolation for any reason. Lack of appropriate contact precautions was particularly common for neonates identified by point prevalence survey as being MRSA-colonized during the first 14 days of ICU stay (only 16% [4 of 25] of MRSA-colonized neonates during the first 14 days were in contact precautions, versus 55% [35 of 64] during days >14, $P<0.001$ for difference, Figure 2).

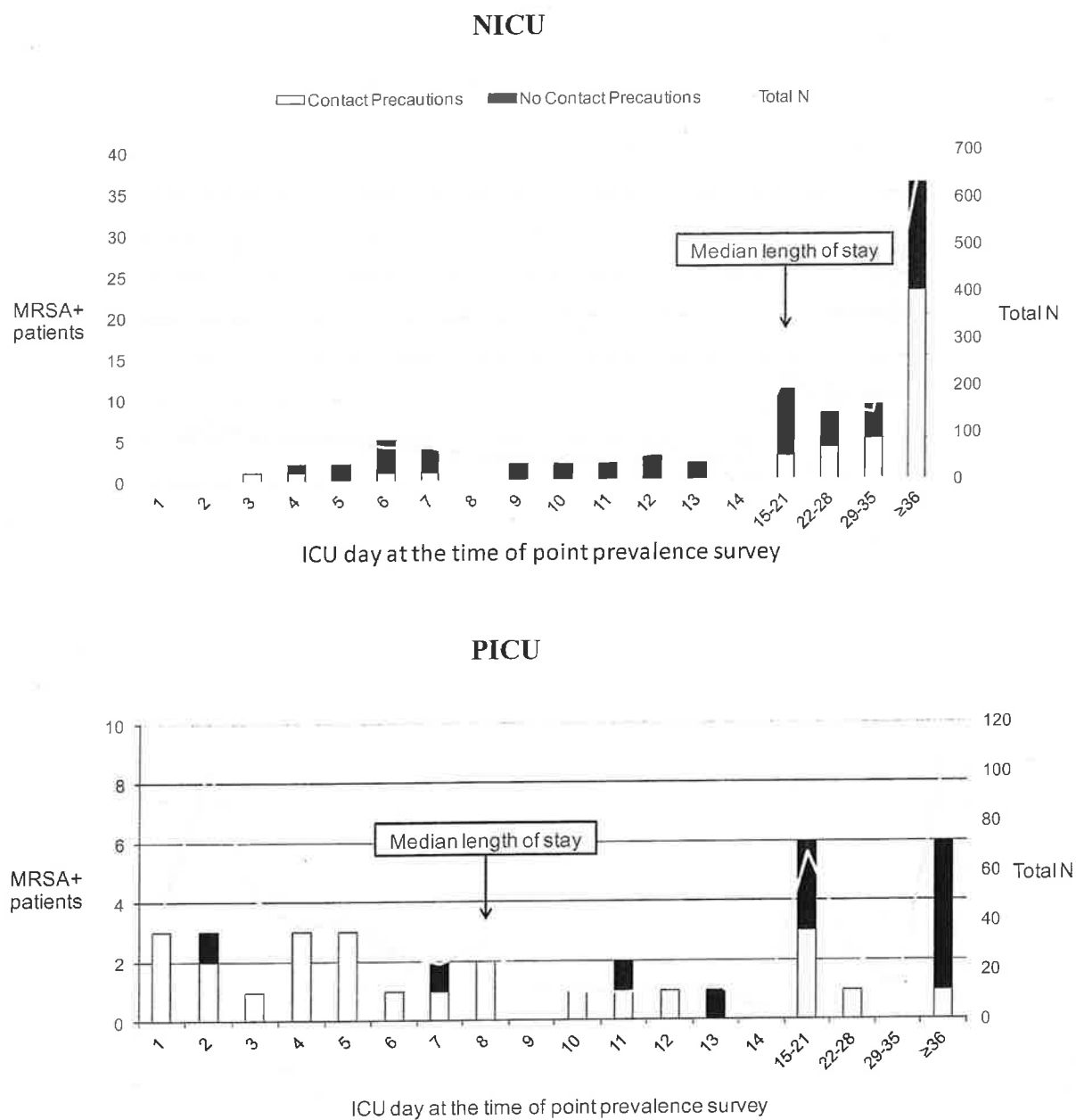


Figure 2. The distribution of MRSA-colonized patients across ICU days of surveillance.

F. Impact of Periodic Active Surveillance in NICUs

In addition to mandated admission screening, 7 of 10 NICUs additionally employed periodic screening strategies (e.g., weekly, twice monthly, or 10 days post-admission) at some point during the 5 year study period, affecting 55% (1162/2101) of participating NICU patients. There was no significant difference in MRSA colonization prevalence during NICU time periods with versus without periodic screening (3.5 vs 5.1% respectively, $P = 0.08$). Furthermore, we did not find a significant difference in rates of appropriate contact precautions during NICU periods with or without periodic screening (42 vs 46%, $P = 0.83$).

G. Performance Characteristics of Testing Different Body Sites for MRSA

Colonization Testing

We assessed the performance characteristics of testing body sites individually for MRSA carriage (nose and umbilicus for NICU patients; nose and groin for PICU patients) using the reference standard of being MRSA positive in any combination of the two body sites during point prevalence testing. For NICU patients who had MRSA culture results available for both nostrils and umbilicus sites, nasal culturing alone identified 87% (62 of 71) MRSA-positive neonates, while 9 patients (13%) were nasal culture negative and umbilicus culture positive for MRSA (Table II). For PICU patients, nasal culturing alone identified 85% (23 of 27) MRSA-positive PICU patients, while 4 patients (15%) were nasal culture negative and groin culture positive for MRSA.

TABLE II

PERFORMANCE CHARACTERISTICS OF INDIVIDUAL
BODY SITE TESTING FOR MRSA COLONIZATION

	Sensitivity % (95% CI)	Negative predictive value % (95% CI)
NICU		
Nose only	87 (77 – 94)	99.4 (99 – 100)
Umbilicus only	55 (43 – 67)	98 (97 – 99)
Nose + umbilicus (Ref)	--	--
PICU		
Nose only	85 (66 – 96)	99 (98 – 100)
Groin only	41 (22 – 61)	97 (95 – 98)
Nose + groin (Ref)	--	--

Note. Since a positive culture for MRSA was always considered a true positive, specificity and positive predictive value = 100% for all body sites.

IV. DISCUSSION

We studied the epidemiology of MRSA colonization among NICU and PICU patients across a spectrum of community and academic hospitals in the city of Chicago, during the 5 years following mandated active surveillance for MRSA. Using serial point prevalence surveys, we identified MRSA colonization in approximately 1 in 25 NICU patients and 1 in 20 PICU patients, with evidence of on-going ICU acquisition in both unit types. We did not find a decrease in MRSA colonization during the 5 years of surveillance; rather, NICU colonization rates were unchanged, and PICU colonization rates increased over time.

This study assessed MRSA epidemiology during a time period in which community-associated MRSA strain types had already become endemic in the United States [13]. Prior single-center studies among NICU patients, performed over prolonged time periods, have found MRSA colonization rates of 1.3 to 1.8% [7, 14]. Higher MRSA colonization rates of 8.6 to 40% have been reported among other single centers during outbreak settings [8, 10, 15]. Among PICU patients, endemic MRSA colonization rates of 3.6 to 4.3% have been reported [16, 17].

The goal of active surveillance is to identify all MRSA-colonized patients to appropriately apply infection control precautions and prevent patient-to-patient transmission. We found high rates of compliance with active surveillance across all NICUs and PICUs. Yet, we found that among MRSA-colonized patients identified through the point prevalence surveys, over half of those in the NICU and a third of those in the PICU were not in contact precautions.

There are several possible explanations for the deficit in appropriate contact precautions: (1) study testing (we used 2 body sites and used broth enrichment) was modestly more sensitive than that used in most hospitals, (2) lag time between hospital admission test collection and result, (3) lag time between positive test result and contact precautions initiation, or (4) MRSA acquisition.

We were not able to directly assess the impact of routine MRSA active surveillance on MRSA prevalence rates, as all hospitals in our study cohort performed mandated MRSA active surveillance; thus, we did not have a non-intervention control group (either historically or concurrently) as a comparator.

This study provides insight into some important knowledge gaps regarding MRSA active surveillance in the pediatric population [18]. Consistent with prior studies [7, 19], we found MRSA to be uncommon among NICU patients within 2 days of admission; therefore, if facilities choose to perform MRSA active surveillance among NICU patients, they should consider performing surveillance at least one additional time point beyond the first 2 ICU days. It is important to note that we did not detect a significant difference in MRSA prevalence or incidence between NICU time periods with and without periodic surveillance; however, lack of difference may have resulted from confounding, as ICUs choosing to perform period surveillance may have been responding to higher MRSA rates.

Our findings also suggest that screening the anterior nares alone is sufficient to detect MRSA colonization. For both the NICU and the PICU populations, nares screening alone had a negative predictive value of 99%, compared to adding the umbilicus site among NICU patients

or groin site among PICU patients. Other studies have supported the nares as the single best site for screening MRSA colonization among NICU patients [8, 20]. Notably, we did not screen the pharynx among PICU patients; this site has been reported in one study to have higher sensitivity compared to nares for MRSA surveillance [17].

Our MRSA surveillance method included testing at 2 body sites as well as broth enrichment of specimens, making the study method more sensitive than that reported by hospitals, potentially biasing estimates of hospital acquisition rates. With respect to extra-nasal testing, the additional MRSA cases recovered was modest (9 additional neonates and 4 additional pediatric patients over the entire study period) and did not substantially change our estimates for ICU acquisition. Furthermore, the primary colonization prevalence outcome, which relied only point prevalence testing results, was unbiased with respect to testing method.

In summary, in a region with mandated MRSA active surveillance, we found on-going MRSA colonization and acquisition in a proportion of NICU and PICU patients across a large metropolitan city. Our findings highlight differences in MRSA epidemiology between NICU and PICU patients, particularly with respect to admission prevalence and timing of acquisition, which may inform future prevention interventions.

CITED LITERATURE

1. Milstone AM, Goldner BW, Ross T, Shepard JW, Carroll KC, Perl TM. Methicillin-resistant *Staphylococcus aureus* colonization and risk of subsequent infection in critically ill children: importance of preventing nosocomial methicillin-resistant *Staphylococcus aureus* transmission. *Clin Infect Dis* **2011**; 53(9): 853-9.
2. Gerber SI, Jones RC, Scott MV, et al. Management of outbreaks of methicillin-resistant *Staphylococcus aureus* infection in the neonatal intensive care unit: a consensus statement. *Infect Control Hosp Epidemiol* **2006**; 27(2): 139-45.
3. Khoury J, Jones M, Grim A, Dunne WM, Jr., Fraser V. Eradication of methicillin-resistant *Staphylococcus aureus* from a neonatal intensive care unit by active surveillance and aggressive infection control measures. *Infection control and hospital epidemiology : the official journal of the Society of Hospital Epidemiologists of America* **2005**; 26(7): 616-21.
4. Nambiar S, Herwaldt LA, Singh N. Outbreak of invasive disease caused by methicillin-resistant *Staphylococcus aureus* in neonates and prevalence in the neonatal intensive care unit. *Pediatric critical care medicine : a journal of the Society of Critical Care Medicine and the World Federation of Pediatric Intensive and Critical Care Societies* **2003**; 4(2): 220-6.
5. Song X, Perencevich E, Campos J, Short BL, Singh N. Clinical and economic impact of methicillin-resistant *Staphylococcus aureus* colonization or infection on neonates in intensive care units. *Infect Control Hosp Epidemiol* **2010**; 31(2): 177-82.
6. Milstone AM, Song X, Beers C, Berkowitz I, Carroll KC, Perl TM. Unrecognized burden of methicillin-resistant *Staphylococcus aureus* and vancomycin-resistant *Enterococcus* carriage in the pediatric intensive care unit. *Infect Control Hosp Epidemiol* **2008**; 29(12): 1174-6.
7. Gregory ML, Eichenwald EC, Puopolo KM. Seven-year experience with a surveillance program to reduce methicillin-resistant *Staphylococcus aureus* colonization in a neonatal intensive care unit. *Pediatrics* **2009**; 123(5): e790-6.
8. Huang YC, Chou YH, Su LH, Lien RI, Lin TY. Methicillin-resistant *Staphylococcus aureus* colonization and its association with infection among infants hospitalized in neonatal intensive care units. *Pediatrics* **2006**; 118(2): 469-74.
9. Murillo JL, Cohen M, Kreiswirth B. Results of nasal screening for methicillin-resistant *Staphylococcus aureus* during a neonatal intensive care unit outbreak. *American journal of perinatology* **2010**; 27(01): 079-81.

10. Kim YH, Chang SS, Kim YS, et al. Clinical outcomes in methicillin-resistant *Staphylococcus aureus*-colonized neonates in the neonatal intensive care unit. *Neonatology* **2006**; 91(4): 241-7.
11. McDougal LK, Steward CD, Killgore GE, Chaitram JM, McAllister SK, Tenover FC. Pulsed-field gel electrophoresis typing of oxacillin-resistant *Staphylococcus aureus* isolates from the United States: establishing a national database. *Journal of clinical microbiology* **2003**; 41(11): 5113-20.
12. Patel JB, Gorwitz RJ, Jernigan JA. Mupirocin resistance. *Clin Infect Dis* **2009**; 49(6): 935-41.
13. David MZ, Daum RS. Update on Epidemiology and Treatment of MRSA Infections in Children. *Current pediatrics reports* **2013**; 1(3): 170-81.
14. Duffy D, Garbush M, Sharland M, Kennea N. Surveillance swabbing for MRSA on neonatal intensive care units—is weekly nasal swabbing the best option? *Journal of Infection Prevention* **2012**; 13(4): 120-4.
15. Carey AJ, Della-Latta P, Huard R, et al. Changes in the molecular epidemiological characteristics of methicillin-resistant *Staphylococcus aureus* in a neonatal intensive care unit. *Infection control and hospital epidemiology : the official journal of the Society of Hospital Epidemiologists of America* **2010**; 31(6): 613-9.
16. Milstone AM, Song X, Beers C, Berkowitz I, Carroll KC, Perl TM. Unrecognized Burden of Methicillin-Resistant *Staphylococcus aureus* and Vancomycin-Resistant *Enterococcus* Carriage in the Pediatric Intensive Care Unit. *Infection Control and Hospital Epidemiology* **2008**; 29(12): 1174-6.
17. Nakamura MM, McAdam AJ, Sandora TJ, Moreira KR, Lee GM. Higher prevalence of pharyngeal than nasal *Staphylococcus aureus* carriage in pediatric intensive care units. *Journal of clinical microbiology* **2010**; 48(8): 2957-9.
18. Milstone AM, Song X, Coffin S, Elward A, Society for Healthcare Epidemiology of America's Pediatric Special Interest G. Identification and eradication of methicillin-resistant *Staphylococcus aureus* colonization in the neonatal intensive care unit: results of a national survey. *Infect Control Hosp Epidemiol* **2010**; 31(7): 766-8.
19. Myers PJ, Marcinak J, David MZ, et al. Universal admission screening for methicillin-resistant *Staphylococcus aureus* in a level IIID neonatal intensive care unit: the first 9 months. *Infection Control and Hospital Epidemiology* **2011**; 32(4): 398-400.
20. Singh K, Gavin PJ, Vescio T, et al. Microbiologic surveillance using nasal cultures alone is sufficient for detection of methicillin-resistant *Staphylococcus aureus* isolates in neonates. *Journal of clinical microbiology* **2003**; 41(6): 2755-7.

VITA

NAME: Rosie Danyell Lyles

EDUCATION: B.S., Biology, Texas Woman's University, Denton, Texas, 1999

M.D., Medical Doctoral, St. Matthew's University School of Medicine, Grand Cayman, 2004

M.H.A., Health Service Administration, St. Joseph's College, Standish, Maine, 2006

M.S., Clinical and Translational Science, University of Illinois, Chicago, Illinois, 2015

PROFESSIONAL EXPERIENCE:

Clinical Affairs Head, Clorox Healthcare Division, July 2014 to Present

Study Director/Co-Investigator: Chicago Epicenter Prevention Programs for the Centers of Disease Control and Prevention (CDC), June 2008 to June 2014

Clinical Quality Assurance Officer/: Infectious Diseases Division at Rush University Medical Center and Cook County Health and Hospital System of Chicago, March 2007 to June 2008

Senior Research Specialist: Infectious Diseases Division at Cook County Health and Hospital System, March 2005- March 2007

PROFESSIONAL MEMEBERSHIP:

American Medical Association
CDC Prevention Epicenters
Infectious Diseases Society of America (IDSA)
The Society for Healthcare Epidemiology of American
Association for Clinical Research Training

PUBLICATIONS:

1. Bleasdale, SC, Trick, WE, Gonzalez, IM, Lyles, RD, Hayden, MK, & Weinstein, RA. Effectiveness of Chlorhexidine Bathing to Reduce Catheter-Associated Bloodstream Infections in Medical Intensive Care Unit Patients. Arch Intern Med 2007; 167(19):2073-9.

2. Schwartz, DN, Wu, U, Lyles, RD, Kieszekowki, P, Xiang, S, Hota, B, & Weinstein, RA. Lost In Translation? Reliability of Assessing Antimicrobial Appropriateness Using Computerized Case Vignettes Assembled from Paper and Computerized Records. Infect Control Hosp Epidemiol 2009; 30(2):163-71.
3. Hota B, Harting B, Weinstein RA, Lyles RD, Bleasdale SC, Trick WE. Electronic algorithmic prediction of central vascular catheter use. Infect Control Hosp Epidemiol 2010; 31:4-11.
4. Bala Hota, Rosie Lyles, Jean Rim, Kyle J. Popovich, Thomas Rice, Alla Aroutcheva, and Robert A. Weinstein, " Predictors of Clinical Virulence in Community-Onset Methicillin-Resistant *Staphylococcus aureus* Infections: The Importance of USA300 and Pneumonia." Clin Infect Dis 2011; 54(8):1-9.
5. Susan S. Huang, MD, MPH; Hilary Placzek, MPH; James Livingston, MBA; Allen Ma, PhD; Fallon Onufrak, BS; Julie Lankiewicz, MPH; Ken Kleinman, ScD; Dale Bratzler, DO; Margaret A. Olsen, PhD, MPH; Rosie Lyles, MD, MHA; Yosef Khan, MD, MPH; Paula Wright, RN, BSN; Deborah S. Yokoe, MD, MPH; Victoria J. Fraser, MD; Robert A. Weinstein, MD; Kurt Stevenson, MD, MPH; David Hooper, MD, "Use of Medicare Claims to Rank Hospitals by Surgical Site Infection Risk following Coronary Artery Bypass Graft Surgery," Infect Control Hosp Epidemiol 2011; 32(8):775-83.
6. Popovich KJ, Lyles R, Hayes R, Hota B, Trick W, Weinstein RA, and Hayden MK. Relationship between chlorhexidine gluconate skin concentration and microbial density on the skin of critically ill patients bathed daily with chlorhexidine gluconate. Infect Control Hosp Epidemiol 2012; 33:889-96.
7. Popovich KJ, Hota B, Aroutcheva A, Kurien L, Patel J, Lyles-Banks R, Grasso A, Spec A, Beavis K, Hayden MK, and Weinstein RA. Community-Associated Methicillin-Resistant *Staphylococcus aureus* Colonization Burden in HIV-Infected Patients. Clin Infect Dis 2013; 56 (8):1067-1074.
8. Michael Y. Lin, Rosie D. Lyles-Banks, Karen Lolans, David W. Hines, Joel B. Spear, Russell Petrak, William E. Trick, Robert A. Weinstein, Mary K. Hayden. The importance of long-term acute care hospitals in the regional epidemiology of *Klebsiella pneumoniae* carbapenemase-producing Enterobacteriaceae. Clinical Infectious Diseases 2013; 57(9):1246-1252.
9. Rosie D. Lyles, Nicholas M Moore, Shayna B. Weiner, Monica Sikka, Michael Y. Lin, Robert A. Weinstein, Mary K. Hayden, Ronda L Sinkowitz-Cochran. Understanding staff perceptions about *Klebsiella pneumoniae* Carbapenemase-producing Enterobacteriaceae control efforts in Chicago long-term acute care hospitals. Infect Control Hosp Epidemiol 2014; 35 (4):367-374.

10. Michael Y. Lin, Karen Lolans, Donald W. Blum, Rosie D. Lyles, Shayna Weiner, Kavya Poluru, Nicholas Moore, David Hines, Robert A. Weinstein, Mary K. Hayden. The effectiveness of routine daily Chlorhexidine Gluconate bathing in reducing *Klebsiella pneumoniae* Carbapenemase-producing *Enterobacteriaceae* skin burden among long-term acute care hospital patients. *Infect Control Hosp Epidemiol* 2014; 35(4):440-442.
11. Michael Klompas , Deverick Anderson , William Trick , Hilary Babcock , Meeta Prasad Kerlin , Lingling Li , Ronda Sinkowitz-Cochran , E Wesley Ely , John Jernigan , Shelley Magill , Rosie Lyles , Caroline O'Neil , Barrett T. Kitch , Ellen Arrington , Michele C. Balas , Ken Kleinman , Christina Bruce , Julie Lankiewicz , Michael V. Murphy , Christopher Cox , Ebbing Lautenbach , Daniel Sexton , Victoria Fraser , Robert A. Weinstein , Richard Platt , and for the CDC Prevention Epicenters. The Preventability of Ventilator-Associated Events: The CDC Prevention Epicenters' Wake Up and Breathe Collaborative. *Am J Respir Crit Care Med*. First published online 04 Nov 2014 as DOI: 10.1164/rccm.201407-1394OC.

ABSTRACTS :

1. Rosie D. Lyles-Banks, Karen Lolans-Mazza, Shayna Weiner, Donald Blom, Nicholas Moore, Michael Y. Lin, Robert A Weinstein, Mary K Hayden. "*Healthcare Worker Hand Carriage of Klebsiella Pneumoniae Carbapenemase-Producing Enterobacteriaceae and other Gram Negative Rods at Long Term Acute Care Hospitals.*" Oral presentation at ID Week, October 2013, San Francisco, California. Abstract #1119.
2. Michael Y. Lin, MD, Donald Blom, Rosie D. Lyles-Banks, Karen Lolans, BS³, Nicholas Moore, Shayna Weiner, Caroline J. Thurlow, Monica K. Sikka, David W. Hines, Robert A Weinstein, Mary K Hayden. *The effectiveness of routine daily chlorhexidine (CHG) bathing in reducing Klebsiella pneumoniae carbapenemase-producing Enterobacteriaceae (KPC) skin burden among long-term acute care hospital (LTACH) patients.* Poster presentation at ID Week, October 2013, San Francisco, California. Abstract #1615.
3. Mary K Hayden, Michael Y. Lin, Rosie D. Lyles-Banks, Shayna Weiner, Nicholas Moore, Karen Lolans, Huiyuan Zhang, Louis Fogg, Donald Blom, Caroline Thurlow, Monica Sikka, David W. Hines, Robert A Weinstein. "*Effect of a Bundled Intervention on Infection and Colonization due to Klebsiella pneumoniae Carbapenemase-Producing Enterobacteriaceae (KPC) at 4 Long-Term Acute Care Hospitals (LTACHs) in Chicago.*" Oral presentation at ID Week, October 2013, San Francisco, California. Abstract #1209.
4. Paul Malpiedi, MPH, Bala Hota, MD, MPH, Shelley Magill, MD, PhD, William Trick, MD, Rosie Lyles, MD, John Martin, MPH, Chris Craver, Scott Fridkin, MD, "*Inter-observer Variability in Bloodstream Infection Determination Using National Healthcare Safety Network Definitions.*" Poster presentation at the 21st Annual Scientific Meeting of the Society for Healthcare Epidemiology of America (SHEA), March 2011, Dallas, Texas.

5. Michael Y. Lin, MD, MPH, Rosie D. Lyles, MD, MHA, Karen Lolans, BS, Mary K. Hayden, MD, Robert A. Weinstein, MD and William E. Trick, MD, "Epidemiology of Methicillin Resistant *Staphylococcus aureus* (MRSA) Among Neonatal Intensive Care Units (ICUs): Implications for Active." Oral Presentation at the 48th Annual Meeting for Infectious Disease Society of America (IDSA), October 2010 Vancouver, Canada. Abstract #2174.
6. Rosie D. Lyles, MD, MHA, Karen Lolans, BS, Mary K. Hayden, MD, Stephen G. Weber, MD, MS, Robert A. Weinstein, MD, William E. Trick, MD, Michael Y. Lin, MD, MPH, "Birds of a Feather? Hospital-level predictors of community-associated versus hospital-associated methicillin-resistant *Staphylococcus aureus* (CA-MRSA vs. HA-MRSA) colonization among intensive care unit (ICU) patient." Poster presentation at the 20th Annual Scientific Meeting of the Society for Healthcare Epidemiology of America (SHEA), March 2010, Atlanta, Georgia.
7. Michael Lin, MD, MPH, Rosie Lyles, MD, MHA, Karen Lolans, B.S., Mary Hayden, MD, Alexander Kallen, MD, MPH, Stephen Weber, MD, MSc, Robert Weinstein, MD and William Trick, MD. "*Glass half empty or half full? The effectiveness of mandated active surveillance in placing methicillin-resistant Staphylococcus aureus (MRSA)-colonized intensive care unit (ICU) patients in Contact Precautions.*" Oral presentation at the 20th Annual Scientific Meeting of the Society for Healthcare Epidemiology of America (SHEA), March 2010, Atlanta, Georgia.
8. Michael Y. Lin, MD, MPH, Rosie D. Lyles, MD, MHA, Karen Lolans, BS, Stephen G. Weber, MD, MS, Craig S. Conover, MD, MPH, Charlesnika T. Evans, MPH, PhD, Mary K. Hayden, MD, Robert A. Weinstein, MD, William E. Trick, MD, "*Region-Wide Epidemiology of Methicillin-Resistant Staphylococcus aureus (MRSA) Among Intensive Care Unit (ICU) Patients Following an Active Surveillance Mandate,*" Oral presentation at the 19th Annual Scientific Meeting of the Society for Healthcare Epidemiology of America (SHEA), March 2009, San Diego, California.
9. Lyles, Rosie, Weinstein, Robert A., Hota, Bala, "*Predictors of Methicillin Resistance in Patients with Serious Staphylococcus aureus Infections,*" Poster presented to 48th Annual ICAAC/IDSA 46th Annual Meeting, October 2008, Washington, D.C. Abstract #L-1498.
10. J.Y. Rim, R.A. Weinstein, R.D. Lyles, B. Hota, "*A Prediction Model for Severe Community-Associated Methicillin-Resistant Staphylococcus aureus Infection (CA-MRSA),*" Oral presentation to 48th Annual ICAAC/IDSA 46th Annual Meeting, October 2008, Washington, D.C.
11. Popovich, KJ, Lyles, RD, Hayes, RA, Hota, B, Weinstein, RA, Hayden, MK, "*Correlation between Results of a Semi-Quantitative (SQ) Chlorhexidine Gluconate (CHG) Indicator Test and Gram-Positive (GP) Colony Counts (CFU) on Skin of MICU Patients (pts) Bathed Daily*

with CHG,” Poster presented at 47th Interscience Conference on Antimicrobial Agents and Chemotherapy (ICAAC), September 2007, Chicago, IL. Abstract #K-1065.

12. David N. Schwartz, MD, R. Scott Evans, M.S, Ph.D., Bernard C. Camins, MD, MSCR, Julie E. Mangino, MD, Monina Klevens, DDS, MPS, Bala Hota, MD, Rosie D. Lyles, MD, James F. Lloyd, BS, Matthew H. Samore, MD, Victoria J. Fraser, MD, Kurt B. Stevenson, MD, MPH, Robert A. Weinstein, MD, “*Electronic measures of Hospital Antimicrobial Utilization: A Multi-Center Pilot Assessment of Feasibility and Variability in Intensive*” Poster presented at the 17th Annual Scientific Meeting of the Society for Healthcare Epidemiology of America (SHEA), April 2007, Baltimore, Maryland. Abstract #36.
13. Schwartz, DN, Lyles, RD, Wu, U, Glowacki, RC, Itokazu, GS, Kieszkowi, P, Xiang, S, Hota, B, Weinstein, RA, “*Computer-Assisted Antimicrobial Recommendations for Optimal Therapy (CAROT): Analysis of Prescribing Errors in a Randomized Trial of Antimicrobial Stewardship Programs*,” Oral presentation at the 17th Annual Scientific Meeting of the Society for Healthcare Epidemiology of America (SHEA), April 2007, Baltimore, Maryland. Abstract #298.
14. Owaisi, AS, Bleasdale, SC, Gonzalez, IM, Lyles, RD, Trick, WE, Hayden, MK, Rice, TW, Aroutchaeva, AA, Weinstein, RA, “*A Descriptive Analysis of Bacteremia (BSIs) in Intensive Care Unit (ICU) Patients Receiving Daily Chlorhexidine (CHG) Skin Cleansing*,” Poster presented to 44th Annual Meeting for Infectious Disease Society of America (IDSA), October 2006, Toronto. Abstract #389.

LECTURES/SEMINARS:

1. Rosie D. Lyles, MD, MHA. “Practical Aspects of Chlorhexidine Bathing in Hospitalized Patients” Meet the Professor Session. Oral presentation at IDweek National Conference, October 11, 2014: 07:00 AM - 08:15 AM, The Pennsylvania Convention Center.
2. Rosie D. Lyles, MD, MHA, et al. “Healthcare Worker Hand Carriage of Klebsiella Pneumoniae Carbapenemase-Producing Enterobacteriaceae and other Gram Negative Rods at Long Term Acute Care Hospitals” Oral presentation at IDweek National Conference, October 4, 2013, San Francisco, CA.
3. Lyles-Banks, Rosie, “Health Careers Opportunity Program,” University of Illinois at Chicago School of Public Health, Summer 2011 and Summer 2012.
4. Lyles, Rosie, “Doctors Back to School Event,” Special Group, Women, Minorities, & GLBT Issues for American Medical Association, 2008 to 2009.
5. **Lyles, RD**, Weinstein, RA, & Vollman, K, “New Evidence-Based Approaches to Preventing Hospital Acquired Infections in the ICU Setting: Emphasis on Resistant Organisms,” Oral

presentation at the National Teaching Institute and Critical Care Exposition, May 2006, Anaheim, CA.

EDUCATIONAL

VIDEO:

Lyles, Rosie, "*Clinical Application of 2% Chlorhexidine Gluconate Cloths for Critically Ill Patients.*" 7 minutes video by Sage Company, Cary, IL. October 2009.